

## Forum Review

# Chemosensitization and Radiosensitization of Tumors by Plant Polyphenols

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### ABSTRACT

The treatment of cancer with chemotherapeutic agents and radiation has two major problems: time-dependent development of tumor resistance to therapy (chemoresistance and radioresistance) and nonspecific toxicity toward normal cells. Many plant-derived polyphenols have been studied intently for their potential chemopreventive properties and are pharmacologically safe. These compounds include genistein, curcumin, resveratrol, silymarin, caffeic acid phenethyl ester, flavopiridol, emodin, green tea polyphenols, piperine, oleandrin, ursolic acid, and betulinic acid. Recent research has suggested that these plant polyphenols might be used to sensitize tumor cells to chemotherapeutic agents and radiation therapy by inhibiting pathways that lead to treatment resistance. These agents have also been found to be protective from therapy-associated toxicities. How these polyphenols protect normal cells and sensitize tumor cells to treatment is discussed in this review. *Antioxid. Redox Signal.* 7, 1630–1647.

### INTRODUCTION

A WIDE VARIETY OF DIETARY phytochemicals have been suggested to block the initiation of cancer or to suppress its development (182). These agents exert their effects by interacting with numerous cellular proteins that, in turn, affect multiple steps in the pathways leading to tumorigenesis. Many of these molecular alterations involve the kinase networks, such as mitogen-activated protein kinases, phosphatidylinositol 3-kinase (PI-3K), and protein kinase, that maintain homeostasis in cells. These kinase pathways converge to activate downstream transcription factors such as nuclear factor- $\kappa$ B (NF- $\kappa$ B) and activator protein 1. In fact, curcumin (turmeric), resveratrol (grapes), genistein (soy), (–)-epigallocatechin gallate (EGCG) (green tea), and other plant phenols are thought to exert their antitumorigenic effects through the inhibition of various mechanisms described in this review (Fig. 1). Furthermore, these plant polyphenols may enhance the tumoricidal effects of chemotherapy and radiotherapy, protect normal cells from therapy-induced damage, and increase systemic bioavailability of chemotherapeutic

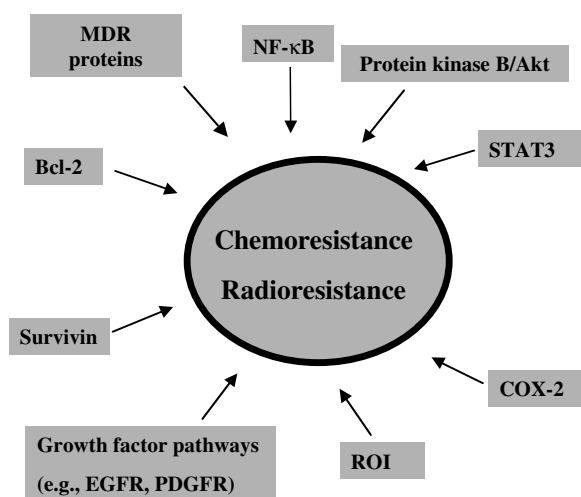
agents (Table 1). However, some of the evidence has come from studies that used superphysiologic doses of these plant polyphenols, and so these effects must be confirmed in clinical trials before the agents can be recommended as safe adjunct treatments.

### MULTIPLE SIGNALING PATHWAYS LEAD TO CHEMORESISTANCE AND RADIORESISTANCE

#### *Reactive oxygen intermediates (ROI) expression*

Potent inducers of both pro-apoptotic and prosurvival pathways, such as members of the tumor necrosis factor (TNF) superfamily, contribute to both the tumor cell death response to anticancer treatments and to the development of resistance to these treatments. These effects are mediated through the production of ROI, which are also known as reactive oxygen species (55). For example, TNF alters the membrane permeability of mitochondria, leading to cytochrome *c*

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**FIG. 1. Cellular molecules implicated in resistance to chemotherapy and radiation therapy.** PDGFR, platelet-derived growth factor receptor.

release and subsequent caspase activation, which, in turn, leads to apoptosis. Furthermore, TNF-associated factor signaling has been shown to be directly linked to the electron transport mechanism in mitochondria and subsequent to ROI production (37). A review placed ROI as a common upstream component in TNF-induced apoptosis as well as in caspase, c-Jun N-terminal kinase, mitogen-activated protein kinase, NF-κB, and activator protein 1 activation (55). Through NF-κB activation, ROI has been shown to mediate prosurvival signaling as well. Therefore, ROI mediates both pro-apoptotic and anti-apoptotic signaling, but the precise mechanisms that lead to these polar outcomes are not yet clear.

Chemotherapy and radiotherapy strongly induce TNF signaling and in doing so use the production of ROI to induce both apoptosis and resistance (Fig. 2). Thought to exert their effects in large part through their antioxidant properties, plant polyphenols may also use ROI in signaling (147).

### *NF-κB activation*

The ubiquitously expressed transcription factor NF-κB is involved in a wide spectrum of cellular responses, including cell cycle control, apoptosis, and stress adaptation. Among the many diseases linked to aberrant NF-κB activation, can-

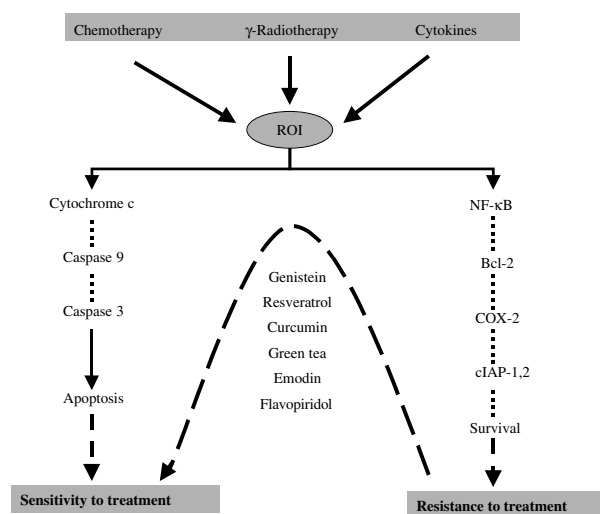
TABLE 1. POLYPHENOLS THAT MAY MODULATE CHEMOTHERAPY AND RADIOTHERAPY

	<i>Polyphenol</i>	<i>Reference</i>
Genistein		
Chemosensitization		
	Enhanced apoptosis induced by docetaxel and cisplatin in pancreatic cancer cells	103
	Enhanced apoptosis induced by doxorubicin, etoposide, or cisplatin in lung cancer cells	95
	Was synergistic with cyclophosphamide in a lung cancer mouse model	205
	Enhanced cisplatin-induced apoptosis in five human melanoma cell lines	188
	Potentiated β-lapachone-induced apoptosis of PC3 and LNCaP prostate cancer cells	92
	Potentiated the effect of dexamethasone on cell cycle progression in liver and colon cancer cells	139
	Enhanced the effects of cisplatin and, to a lesser extent, of vincristine in meduloblastoma cells	84
	Enhanced the effects of tiazofurin in human leukemia cells	100
	Enhanced the effects of tiazofurin in human ovarian carcinoma cells	99
	Was synergistic with quercetin in ovarian carcinoma cells	169
Radiosensitization		
	Potentiated the effect of radiation on prostate carcinoma cells	65
	Improved the efficacy of chemoradiotherapy in a murine lung tumor model	111
	Enhanced radiosensitivity in human esophageal cancer cell lines	4
	Enhanced γ-irradiation-induced apoptosis and cell cycle arrest in K562 leukemia cells	138
Resveratrol		
Chemosensitization		
	Enhanced paclitaxel-induced apoptosis of lung cancer cells	91
	Reduced paclitaxel-induced apoptosis in human neuroblastoma SH-SY5Y cells	130
	Reduced TNF-induced apoptosis in human leukemia cells	109
	Reduced H <sub>2</sub> O <sub>2</sub> -induced apoptosis in human neuronal PC12 cells	73
	Inhibited H <sub>2</sub> O <sub>2</sub> , vincristine-, daunorubicin-, and C2-induced apoptosis in human leukemia cells	2
	Sensitized multiple myeloma cells and non-Hodgkin's lymphoma cells to paclitaxel-induced apoptosis	25
	Reduced H <sub>2</sub> O <sub>2</sub> -induced apoptosis in leukemia cells by inhibiting leukotriene B4 and prostaglandin E2	108
Radiosensitization		
	Enhanced γ-radiation-induced apoptosis and cell cycle arrest of Hela and SiHa cells	216
Curcumin		
Chemosensitization		
	Sensitized multiple myeloma cells to vincristine and melphalan	25
	Potentiated the cytotoxic effects of doxorubicin, 5-FU, and paclitaxel in prostate cancer cells	67

(Continued)

TABLE 1. CONTINUED

<i>Polyphenol</i>	<i>Reference</i>
Radiosensitization	
Sensitized PC3 prostate cancer cells to radiation through suppression of TNF production	41
Protective effects	
Reduced lung toxicity in rats treated with whole-body radiation	191
Reduced radiation-induced genotoxicity in mice treated with whole-body radiation	192
Decreased acute toxicity from whole-body radiation in rats	71
Protected rats from doxorubicin-induced nephrotoxicity	200
Protected rats from doxorubicin-induced cardiotoxicity	199
Reduced mucosal injury from trinitrobenzene sulfonic acid-induced colitis in mice	196
Inhibited camptothecin-, mechlorethamine-, and doxorubicin-induced apoptosis of breast cancer cells	176
Green tea polyphenols	
Chemosensitization	
Enhanced antitumor activities of other anthracyclines, cisplatin, and irinotecan in ovarian sarcoma-bearing mice	180
Enhanced antitumor effect of doxorubicin in mice with ascites carcinoma	158
Enhanced inhibitory effects of doxorubicin against Ehrlich ascites carcinoma in mice (green tea)	159
Was synergistic with doxorubicin in mice with M5076 ovarian sarcoma	179
Was synergistic with doxorubicin in inhibition of hepatic metastasis of M5076 ovarian sarcoma in mice	158, 178, 179
Reversed MDR and increased doxorubicin concentration in mice with P388 leukemia	160
Increased pirarubicin concentration in M5076 ovarian sarcoma cells	181
Increased antitumor activity of idarubicin and reduced its toxicity in mice with P388 leukemia	161
Emodin	
Chemosensitization	
Enhanced arsenic trioxide-induced apoptosis in HeLa cells	208
Potentiated effects of <i>cis</i> -platinol, doxorubicin, and 5-FU in Merkel cell carcinoma cells (aloe-emodin)	52
Enhanced effects of paclitaxel on Her-2/neu-overexpressing breast cancer cells	213
Enhanced cytotoxic effects of cisplatin, doxorubicin, and etoposide on Her-2/neu-overexpressing non-small cell lung cancer cells	210
CAPE	
Protective effects	
Protected rats from doxorubicin-induced cardiotoxicity	51
Protected rats from cisplatin-induced nephrotoxicity	135
Protected rats from bleomycin-induced lung fibrosis	136
Protected rats from ischemia-reperfusion kidney injury	61
Protected rats from radiation-induced inflammatory changes	106
Flavopiridol	
Chemosensitization	
Potentiated effects of mitomycin c in breast and gastric cancer cells	165
Was synergistic with paclitaxel, cytarabine, topotecan, doxorubicin, etoposide, and cisplatin in non-small cell lung cancer cells	29
Enhanced doxorubicin-induced apoptosis in retinoblastoma protein-deficient sarcoma cells	101
Augmented CPT-11-induced apoptosis in Hct116 colon cancer cells	123
Increased sensitization to gemcitabine in gastrointestinal cancer cells	77
Radiosensitization	
Potentiated $\gamma$ -radiation-induced apoptosis in colon and gastric cancer cells	78
Enhanced radiosensitivity of ovarian carcinoma cells	150
Silymarin	
Chemosensitization	
Potentiated doxorubicin cytotoxicity in P-glycoprotein-positive breast cancer cells (silymarin)	214
Enhanced doxorubicin-induced growth inhibition in human prostate cancer cells (silymarin)	195
Was synergistic with cisplatin and doxorubicin in breast cancer and ovarian carcinoma cells (silybin)	164
Protective effects	
Protected rat cardiomyocytes from anthracycline-induced toxicity	42
May have protected rat heart membrane from doxorubicin-induced damage	148
Ginger	
Protective effects	
Protected rats and dogs from cisplatin-induced nausea	167, 168
Piperine	
Effects on bioavailability	
Increased bioavailability of propranolol and theophylline in humans	15
Enhanced bioavailability of aflatoxin B1 in rat tissues	6
Enhanced bioavailability of curcumin in rats and humans	173



**FIG. 2.** Chemotherapy and radiotherapy induce both pro-apoptotic signaling, leading to cell death and treatment sensitivity, and anti-apoptotic signaling, leading to increased cell survival and treatment resistance. Plant polyphenols may help increase sensitivity and inhibit the pathways leading to resistance.

cer has been the major focus because of NF- $\kappa$ B's role as a central regulator of the inflammatory response, its regulation of genes involved in cellular survival (Bcl-2, Bcl-xL, c and x forms of inhibitor of apoptosis protein [IAP], and superoxide dismutase) and tumor progression (intercellular adhesion molecule 1, vascular cell adhesion molecule 1, endothelial leukocyte adhesion molecule 1, cyclooxygenase-2 [COX-2], inducible nitric oxide synthase [iNOS], and matrix metalloproteinase [MMP-9]). Furthermore, its constitutive activity has been frequently elevated in many types of tumors, including leukemia, lymphoma, prostate cancer, breast cancer, colon cancer, melanoma, and head and neck cancers (54, 152). Ursolic acid and betulinic acid have been shown to exert their effects through mechanisms that involve the inhibition of NF- $\kappa$ B (172, 186). Also, NF- $\kappa$ B is activated by chemotherapeutic agents and ionizing radiation, and can lead to treatment-induced tumor cell resistance (45, 56, 79). Furthermore, the suppression of NF- $\kappa$ B by different methods can sensitize tumor cells to both chemotherapy and radiotherapy.

*In vitro* and *in vivo* studies have shown that constitutive activation of NF- $\kappa$ B inhibits chemotherapy-induced apoptosis in a number of tumor types (23, 24, 202). For example, up-regulation of NF- $\kappa$ B-inducible genes protected MDA-MB-231 breast cancer cells from paclitaxel-induced and radiation-induced apoptosis (129, 202), and chemotherapy-induced NF- $\kappa$ B activation led resistance to conventional cancer treatment resistance (18, 20, 137, 202). In a recent study of four tumor cell lines, each treated with different chemotherapy regimens (doxorubicin, 5-fluorouracil [5-FU], cisplatin, and paclitaxel), cell survival correlated with the level of NF- $\kappa$ B activity induced by the drugs (43). In another study, NF- $\kappa$ B-binding activity decreased in breast cancer cells treated with anti-Her-2/neu antibody (Trastuzumab; Herceptin), suggesting a role for NF- $\kappa$ B in the therapeutic ef-

ficacy of this antibody combined with chemotherapy for patients with Her-2/neu-positive breast cancer (144). Others studies have shown tumor regression due to tumor sensitization to CPT-11 (irinotecan), a topoisomerase I inhibitor, in mouse xenograft models that overexpress inhibitory subunit of NF- $\kappa$ B (I $\kappa$ B $\alpha$ ) (45, 203). Additional reports using adenovirus-mediated gene transfection of I $\kappa$ B $\alpha$  demonstrated the potentiation of chemotherapeutic efficacy in both *in vitro* and *in vivo* models of gastrointestinal malignancies (46, 203), human glioma cells (204), and pancreatic cancer cells (11). Compared with cells without the super-repressor, fibrosarcoma cells expressing the I $\kappa$ B $\alpha$  super-repressor were more sensitive to radiation-induced apoptosis (17), and glioblastoma cell lines (A172, M054) expressing the super-repressor were more susceptible to radiotherapy (206). Protease inhibitors are less specific but are a more clinically useful way to examine the effects of NF- $\kappa$ B inhibition. In a preclinical study, the use of protease inhibitors increased radiation-induced apoptosis of lymphoma cells (93).

### COX-2 expression

COX-2, an enzyme expressed largely in response to inflammatory disorders and cancer, mediates prostaglandin production and is currently under clinical investigation as a target for anticancer therapy (120, 125). Its presence has been associated with more aggressive tumor phenotypes and worse outcome for patients with breast cancer, colon cancer, head and neck cancers, lung cancer, and pancreatic cancer. Accumulating evidence suggest that COX-2 is involved in multiple aspects of carcinogenesis, including maintenance of tumor growth, facilitation of metastatic spread, and resistance to various therapies (89, 116). As a result, selective inhibitors of COX-2, such as SC-236 and celecoxib, have been used *in vitro* and *in vivo* to test whether its inhibition may sensitize tumor cells to chemotherapy and radiotherapy. For example, SC-236 increased tumor radioresponse in murine tumor models and in a human glioma xenograft in nude mice (87, 118, 142), and celecoxib enhanced the response of A431 human tumor xenografts in nude mice to radiation and docetaxel chemotherapy, radiotherapy, or both (188). The mechanisms that account for these findings and others are not yet clear but may involve the elimination of prostaglandins as protective molecules in response to chemotherapy and radiotherapy (117).

### Bcl-2 expression

Bcl-2 is a member of the Bcl-2 family of proteins that regulates both pro-apoptotic and anti-apoptotic signaling in cells. Bcl-2 itself is an anti-apoptotic protein that is inappropriately overexpressed in a number of solid and hematopoietic tumors, and exerts its influence by enhancing cellular survival (81), which contributes to resistance to conventional treatments, including chemotherapy and radiotherapy (36). Furthermore, several investigations have shown that inhibiting Bcl-2 sensitizes tumor cells to chemotherapy and radiotherapy. For example, transfection with the gene *PTEN*, which down-regulates Bcl-2, potentiated the effects of radiotherapy in several types of prostate cancer cells (156). An-

other study, using Bcl-2 antisense oligonucleotide, demonstrated increased apoptosis and enhanced chemotherapeutic efficacy in undifferentiated thyroid carcinomas (86). These results and others (21, 62) have highlighted Bcl-2 as a potential target for chemosensitization and radiosensitization.

### *Survivin expression*

Survivin is a member of the mammalian IAP family and functions primarily to inhibit the apoptosis pathway by blocking caspase 9 activation (162, 170). Both *in vitro* and *in vivo* experiments have demonstrated its cancer-inducing properties (16) and its overexpression in several malignant tissues and its absence in most normal tissues (7, 8). Survivin has been shown to be involved in angiogenesis and to be necessary for the anti-apoptotic effect of vascular endothelial growth factor (114, 132, 193, 215). Furthermore, high levels of survivin have been associated with a high rate of tumor recurrence, low overall patient survival, and high tumor resistance to chemotherapy and radiotherapy (7). Cell sensitization to therapies has been demonstrated in several studies: Inhibition of survivin with a ribozyme or dominant-negative molecule sensitized melanoma and pancreatic cancer cells to radiation (12, 141); survivin inhibition sensitized breast cancer cells to paclitaxel and doxorubicin (133, 201); and survivin inhibition plus radiotherapy resulted in significantly decreased survival of lung cancer cells (107).

### *Multidrug resistance (MDR) protein expression*

MDR proteins and MDR-associated proteins are two subfamilies of the ATP-binding cassette family proteins that regulate the P-glycoproteins that remove drugs from cells at the cost of ATP hydrolysis (49, 166). The relationship between MDR protein expression and chemoresistance is fairly well established (96, 185). For example, *in vitro* and *in vivo* investigations have suggested the involvement of MDR proteins in multiple mechanisms that lead to chemoresistance in prostate cancer (197). This result was consistent with that from a study that used an NF- $\kappa$ B inhibitor to down-regulate the expression of MDR proteins, thereby leading to increased apoptosis in prostate cancer cells (53). The association between MDR protein expression and radioresistance is less clear. For example, cancer cells that expressed the *MDR* gene were no more resistant to radiotherapy than their non-MDR counterparts (57).

### *AKT expression*

The protein kinase B/Akt pathway is a downstream effector of PI-3K and has been described as a mediator of anti-apoptotic signaling in cancer cells. Akt has been shown to affect cell cycle progression and foster tumorigenesis when overexpressed (209). This overexpression may contribute to chemoresistance and radioresistance. The ectopic expression of constitutively active Akt1 resulted in enhanced resistance of non-small cell lung cancer cells to a panel of chemotherapeutic agents (68). In breast cancer cells, inhibition of the PI-3K/Akt pathway led to enhanced paclitaxel, doxorubicin, and 5-FU cytotoxicity (75), and dominant-negative expres-

sion vectors sensitized the cells to the induction of apoptosis by paclitaxel, doxorubicin, 5-FU, etoposide, and camptothecin (88). However, in pancreatic cancer cells, the PI-3K/Akt pathway did not seem to be involved in gemcitabine resistance (11). Accumulating evidence has suggested that the PI-3K/Akt pathway may also be a major contributor to radioresistance (209). One report indicated that activated Akt in bile duct cancer cells was associated with radioresistance, as evidenced by the enhanced radiosensitization via Akt inhibition (189). This report and another (112) indicated the PI-3K/Akt pathway as a target for tumor sensitization to radiotherapy.

### *Signal transducer and activator transcription (STAT)3 expression*

STAT3 is a ubiquitously expressed member of the STAT family of transcription factors that is activated by tyrosine phosphorylation via upstream receptors that bind to growth factors such as epidermal growth factor (EGF) and platelet-derived growth factor and cytokines such as interleukin-6 (IL-6) (5, 97, 98). STAT3 has diverse biologic functions, including cell growth regulation, apoptosis regulation, and cell differentiation. STAT3 has also been shown to be constitutively active in a number of human cancers and to be necessary for tumor cell growth (32). Moreover, STAT3 may mediate chemoresistance, and its inhibition may sensitize cells to apoptosis. For example, STAT3 inhibition contributed to the decreased survival of multiple myeloma cells (26), and it may have sensitized pancreatic cancer cells to apoptosis (59). Blockade of STAT3 using various techniques sensitized estrogen receptor-negative breast cancer cells to chemotherapy-induced apoptosis (153). Finally, STAT3 inhibition with a STAT3 antisense oligonucleotide enhanced radiation-induced apoptosis in prostate cancer cells (36).

### *EGF receptor (EGFR) expression*

EGFR is a transmembrane glycoprotein with intrinsic tyrosine kinase activity. On binding with EGF or transforming growth factor-beta, EGFR regulates a signaling cascade that, in turn, regulates cell growth and proliferation. EGFR overexpression has been linked to aggressive tumor phenotypes, poor patient prognosis, and poor tumor response to therapies (113). For example, the magnitude of EGFR expression correlated with increased tumor chemoresistance and radioresistance in a variety of *in vivo* tumors, including murine carcinoma, squamous cell carcinoma, ovarian adenocarcinoma, hepatocarcinoma, and adenosquamous carcinoma (3, 119, 126). Use of the anti-EGFR antibody C225 greatly enhanced the response of A431 human tumor xenografts in nude mice to docetaxel, radiation, or both (119, 126). Other investigators also reported the increased sensitization of tumor cells to radiotherapy through EGFR inhibition (28, 69, 70). Mechanisms that account for this effect by EGFR inhibitors are becoming clearer with time, and likely involve sensitization to treatment-induced apoptosis, inhibition of treatment-induced repair mechanisms, and inhibition of tumor angiogenesis (120).



## BACKGROUND ON SELECTED POLYPHENOLS

Genistein is a soy-derived isoflavone that acts as a tyrosine kinase inhibitor and has a structure with an affinity for the estrogen receptor and androgen-mediated pathways (33). Therefore, it is not surprising that it has received considerable attention as a chemopreventive agent in breast, prostate, and other cancers (163). In fact, epidemiologic studies have linked the lower incidences of breast and prostate cancers in Asian populations than in non-Asian populations to significant differences in diet, including a higher concentration of soy isoflavones such as genistein. Genistein is thought to inhibit the growth of cancer cells by modulating genes related to cell cycle control and apoptosis and is a potent inhibitor of angiogenesis and metastasis. *In vitro* and *in vivo* investigations have shown that this mechanism may be mediated through NF- $\kappa$ B inhibition (128). Furthermore, genistein combined with chemotherapy and radiotherapy may enhance the efficacy of these therapies.

Resveratrol (*trans*-3,5,4'-trihydroxystilbene) was first isolated in 1940 as a constituent of the roots of white hellebore (*Veratrum grandiflorum* O. Loes) but has since been found in various plants, including grapes, berries, and peanuts. Research in recent years has focused on the anticancer properties of resveratrol, as suggested by its ability to suppress the proliferation of lymphoid and myeloid cancers, multiple myeloma, breast cancer, prostate cancer, colon cancer, pancreatic cancer, melanoma, head and neck squamous cell carcinoma, ovarian carcinoma, and cervical carcinoma (14, 35, 50).

Curcumin, a diferuloylmethane derived from turmeric (*Curcuma longa*), has been the subject of intense study as a chemopreventive agent and as a complement to chemotherapy and radiotherapy. Notable findings in recent years have included its ability to suppress proliferation in a variety of tumor cell types; down-regulate NF- $\kappa$ B target genes such as *COX-2*, *iNOS*, *MMP-9*, *urokinase-type plasminogen activator*, and *cyclin D1*; inhibit the expression of growth factor receptors, including EGFR and human EGFR 2; and inhibit several protein kinases involved in the signaling pathways leading to tumorigenesis (1). Moreover, curcumin inhibits NF- $\kappa$ B activation induced by various inflammatory stimuli (54, 124, 151), the I $\kappa$ B kinase (IKK) activation needed for NF- $\kappa$ B activation (17, 31, 122), and NF- $\kappa$ B-induced osteoclastogenesis (27). An epidemiologic study suggested that low incidences of gastrointestinal malignancies in India may be attributable to the presence of natural additives, including curcumin, in the Indian diet (121).

Used as a medicinal agent in Asia for more than 4,000 years, green tea is derived from the leaves of *Camellia sinensis* and is an efficacious chemopreventive agent and modulator of chemotherapy (140, 182). Several investigations, including a phase I clinical trial (146), have demonstrated a decreased relative risk of many cancers, including lung, colorectal, pancreatic, and stomach cancers with the consumption of green tea. Growing evidence from animal studies has also suggested the chemopreventive potential of green tea (140). Two components of green tea, EGCG and  $\gamma$ -glutamylethylamide (theanine), have been the particular

focus of ongoing research. Several other plant polyphenols (e.g., emodin, caffeic acid phenethyl ester [CAPE], flavopiridol, silymarin, ginger, and piperine) are currently under investigation and are discussed below.

## SENSITIZATION OF TUMOR CELLS BY POLYPHENOLS TO CHEMOTHERAPEUTIC AGENTS

### Green tea polyphenols

Mounting evidence has suggested that the amino acid theanine, a major component of green tea, plus chemotherapy has a synergistic effect for a variety of cancers. An investigation of the combined effects of theanine and glutamate transporter inhibitors on the antitumor activity of doxorubicin in M5076 ovarian sarcoma-bearing mice revealed that, compared to the doxorubicin-alone group, theanine significantly enhanced the inhibitory effect of doxorubicin on tumor growth and increased the drug's concentration in the tumors (188). Oral administration of theanine or green tea similarly enhanced the antitumor activity of doxorubicin. Theanine plus doxorubicin also suppressed the hepatic metastasis of ovarian sarcoma. An increase in doxorubicin concentration was not observed in normal tissues such as the liver and heart. The investigators described novel mechanisms of enhancement of antitumor efficacy of doxorubicin via the inhibition of glutamate transporters by theanine. Moreover, theanine enhanced the antitumor activities of other anthracyclines, cisplatin, and irinotecan (188).

Another study showed that, compared with doxorubicin alone, theanine plus doxorubicin enhanced the antitumor effect over twofold in mice with Ehrlich ascites carcinoma (158). Furthermore, theanine caused an almost threefold increase in the concentration of doxorubicin within the tumor, which correlated with the increased efficacy of the drug combination and decreased tumor weight (188). Consistent with these results, theanine and doxorubicin acted synergistically in mice with M5076 ovarian sarcoma, enhancing tumor concentration of chemotherapy, inhibiting tumor growth, and decreasing hepatic metastasis (159, 178, 179). Theanine was also effective in reversing MDR in mice with P388 leukemia by increasing the doxorubicin concentration within the tumor and decreasing tumor weight (160). In both the ovarian sarcoma and leukemia cells, theanine attacked the same transport process for doxorubicin, elevated the doxorubicin concentration, and increased the doxorubicin-induced antitumor activity. Other studies have also demonstrated the inhibition of MDR-associated proteins by green tea components (44, 66).

Like theanine, the green tea components—caffeine, EGCG, and flavonoids—inhibit doxorubicin efflux from Ehrlich ascites carcinoma cells. Thus, EGCG and flavonoids may enhance doxorubicin-induced antitumor activity and increase doxorubicin concentration in tumors by inhibiting the efflux. These components in green tea probably exhibit low toxicity, and green tea plus chemotherapy probably has few adverse effects. Similar synergistic effects have been noted

for theanine plus pirarubicin (181, 194), idarubicin (161), cisplatin (155, 217), and CPT-11 (175). However, one report found no association between intake of green tea and risk of breast cancer in Japanese women (183).

Studies have demonstrated the potent chemopreventive effect of EGCG in several types of cancer cells. In the prostate cancer cell lines LNCaP and DU-145, EGCG inhibited NF- $\kappa$ B activation, thereby leading to increased apoptosis and decreased expression of the prometastatic genes encoding for MMP-2 and MMP-9 (60, 63, 198). In human epidermoid carcinoma A431 cells, EGCG activated caspases, which led to increased apoptosis and inhibition of NF- $\kappa$ B (60). In YCU-H891 head and neck cancer cells and MDA-MB-231 breast carcinoma cell lines, EGCG inhibited both constitutive NF- $\kappa$ B activation and EGFR activation and strongly inhibited vascular endothelial growth factor production (110). Also in breast cancer cells, EGCG inhibited basal Her-2/neu receptor tyrosine phosphorylation, leading to subsequent inhibition of PI-3K, Akt, and NF- $\kappa$ B activation (145). In the lung cancer cell line PC-9, EGCG inhibited NF- $\kappa$ B-inducing kinase, an upstream kinase leading to I $\kappa$ B kinase and NF- $\kappa$ B activation (134). Finally, in EGF- and 12-*O*-tetradecanoylphorbol 13-acetate-stimulated mouse JB6 epidermal cells, EGCG inhibited ultraviolet B-induced NF- $\kappa$ B-mediated transcription (131). Taken together, these results suggested that the chemopreventive properties of EGCG may derive from the modulation and inhibition of a variety of signaling pathways that lead to NF- $\kappa$ B activation and tumorigenesis.

### Genistein

Several reports have highlighted the enhanced efficacy of chemotherapy when it is combined with several plant polyphenols. Genistein has been shown to potentiate the effects of chemotherapy for numerous tumor types. In the pancreatic cancer cell line, treatment with genistein before docetaxel or cisplatin administration enhanced tumor cell death compared with treatment with either chemotherapeutic drug alone. This effect may have been mediated by the inhibition of NF- $\kappa$ B by genistein, causing increased apoptosis (188). In EGFR-expressing lung cancer cells, genistein combined with cisplatin, doxorubicin, or etoposide enhanced the antiproliferative effects of these drugs and induced programmed cell death (95). This result was consistent with that from another report using a lung cancer mouse model to show the synergistic effect of genistein and cyclophosphamide (205). In another study using five human melanoma cell lines, genistein plus cisplatin enhanced apoptosis in all cell lines versus either treatment alone; furthermore, genistein significantly reduced levels of the anti-apoptotic proteins Bcl-2 and Bcl-xL and increased levels of the pro-apoptotic protein Apaf-1 (188). In prostate cancer cell lines PC3 and LNCaP, genistein plus  $\beta$ -lapachone resulted in more potent cell killing than either treatment alone did (92). In liver and colon cancer cell lines, genistein plus dexamethasone resulted in enhanced expression of p21<sup>WAF1/CIP1</sup>, an inhibitor of cyclin-dependent kinase 2, thereby halting cell cycle progression (139). In medulloblastoma cells, genistein potentiated the effects of cisplatin and, to a lesser extent, those of vincristine (84). In human leukemia cells (188) and ovarian carcinoma cells (188), genis-

tein plus tiazofurin resulted in greater cells growth inhibition and increased cell differentiation compared to either treatment alone. Finally, genistein and quercetin acted synergistically in ovarian carcinoma cells (169).

The chemopreventive outcomes of genistein in several tumor types may be achieved by targeting an NF- $\kappa$ B-dependent pathway. For example, in studies using MDA-MB-231 breast cancer cells and PC3 prostate cancer cells, genistein inhibited cell growth and induced apoptosis via the down-regulation of Akt and NF- $\kappa$ B pathways (58, 102). In T-cell lymphoma cell lines, genistein increased pro-apoptotic caspase 3 activity and reduced activity of the NF- $\kappa$ B-mediated anti-apoptotic factors A1 and cIAP-1 (19). These observations were consistent with those from an *in vivo* investigation of pancreatic cancer, in which genistein significantly improved patient survival, almost completely inhibited metastasis, and increased apoptosis by activating caspase 3 (34). Other studies have shown that genistein inhibited NF- $\kappa$ B-binding activity in human hepatocarcinoma cells (184), prostate cancer cells LNCaP and PC3 (47), and human alveolar epithelial carcinoma cells (38).

### Resveratrol

Resveratrol has been shown to potentiate the apoptotic effects of cytokines, chemotherapeutic agents, and  $\gamma$ -radiation. One study assessed the *in vitro* biologic activity of resveratrol in lung cancer cell lines by examining its effect on apoptosis induced by paclitaxel (91). Simultaneous exposure to resveratrol and paclitaxel did not result in significant synergy, but pretreatment with resveratrol (10  $\mu$ M for 3 days) significantly enhanced the subsequent antiproliferative effect of paclitaxel. The study also examined the effects of resveratrol and paclitaxel on p21<sup>WAF1</sup>, p27<sup>KIP1</sup>, E-cadherin, EGFR, and Bcl-2 levels in non-small cell lung cancer EBC-1 cells. Resveratrol (10  $\mu$ M for 3 days) administered before paclitaxel increased p21<sup>WAF1</sup> expression approximately fourfold. These results suggested that lung cancer cells exposed to resveratrol have a lowered threshold for killing by paclitaxel and, thus, that resveratrol may be a compromising alternative therapy for lung cancer (91).

Another study found that resveratrol modified the expression of apoptotic regulatory proteins and sensitized non-Hodgkin's lymphoma and multiple myeloma cell lines to paclitaxel-induced apoptosis (74). Resveratrol down-regulated the expression of anti-apoptotic proteins Bcl-xL and myeloid cell differentiation factor 1 and up-regulated the pro-apoptotic proteins Bax and Apaf-1. Furthermore, inhibition of *Bcl-xL* (an NF- $\kappa$ B target gene) by resveratrol was critical for chemosensitization because functional impairment of Bcl-xL mimicked resveratrol-mediated sensitization to paclitaxel-induced apoptosis (91). In contrast, another study found that *trans*-resveratrol reduced cellular death in SH-SY5Y neuroblastoma cells exposed to paclitaxel by inhibiting paclitaxel-induced activation of caspase 7 and the degradation of poly(ADP-ribose) polymerase (91).

The contrasting effects of resveratrol may be dose-dependent, whereby it potentiates the effects of cytokines and chemotherapeutic agents at higher concentrations and inhibits their effects at lower concentrations. At relatively low con-

centrations, resveratrol abrogated TNF-induced cytotoxicity and caspase activation (109) and attenuated  $H_2O_2$ -induced cytotoxicity, DNA fragmentation, and intracellular accumulation of ROI (73). Furthermore, a recent report showed that low concentrations of resveratrol (4–8  $\mu M$ ) inhibited caspase activation, DNA fragmentation, and translocation of cytochrome *c* induced by  $H_2O_2$ , vincristine, daunorubicin, etc. (2). The effects of resveratrol at these levels were attributed to increased intracellular superoxide concentration and reduced drug-induced acidification. These results suggested that the protective or inhibitory effects of resveratrol at low concentrations are secondary to its antioxidant mechanism.

### Curcumin

Several studies have shown that curcumin sensitizes cells to chemotherapy. One report showed that curcumin down-regulated NF- $\kappa$ B and sensitized multiple myeloma cells to vincristine and melphalan. Furthermore, the NF- $\kappa$ B target genes *Bcl-2*, *Bcl-xL*, *cyclin D1*, and *IL-6* were down-regulated by curcumin, leading to the suppression of proliferation and arrest of cells at the G<sub>1</sub>/S phase of the cell cycle (25). In another study, curcumin potentiated the cytotoxic effects of doxorubicin, 5-FU, and paclitaxel in prostate cancer cells, and suppressed both the constitutive and TNF-induced activation of NF- $\kappa$ B (67). Likewise, doxorubicin-induced NF- $\kappa$ B activation was attenuated by curcumin (43). Curcumin has also been shown to modulate the activity of the MDR genes, thereby suppressing drug efflux by P-glycoprotein and leading to chemosensitization (10, 105). These results, when taken together, support the further investigation of curcumin plus chemotherapeutic drugs as a synergistic therapy.

### Emodin

Emodin may sensitize cancer cells to chemotherapy. For example, it sensitized HeLa cells to arsenic trioxide via generation of ROI and ROI-mediated inhibition of two major prosurvival transcription factors, NF- $\kappa$ B and activator protein 1 (208), and it effected apoptosis in human promyelocytic HL-60 cells by inducing caspase 3, but independently of ROI (40). In a study using several chemotherapeutic agents including *cis*-platinol (abiplastin), doxorubicin (adriablastin), 5-FU, and tyrosine kinase inhibitor STI 571, aloe-emodin potentiated their inhibitory effects on Merkel cell carcinoma cells (52). In cholangiocarcinoma cells and epithelial stem-like cells, emodin and celecoxib synergistically suppressed cell growth and increased activation of caspases 9 and 3; this mechanism was suggested to work through Akt inhibition (94). In Her-2/neu-overexpressing breast cancer cells, emodin inhibited Her2/neu tyrosine kinase activity, suppressed cell growth, induced cell differentiation, and sensitized cells to paclitaxel chemotherapy (211, 212, 213). In Her-2/neu-overexpressing non-small cell lung cancer cells, emodin suppressed cell proliferation, reduced Her-2/neu tyrosine kinase activity, and enhanced the inhibitory effects of cisplatin, doxorubicin, and etoposide on cell proliferation (210). However, one study showed no chemosensitizing effect of emodin in ovarian cancer cells (64).

### Flavopiridol

Flavopiridol potentiates the effects of chemotherapy in several types of cancer. For example, flavopiridol potentiated the cytotoxic effects of mitomycin c by promoting drug-induced apoptosis in breast and gastric cancer cells; this effect was dependent on the order of treatment (165). An investigation using non-small cell lung cancer cells revealed that the sequence of administration was important to the sensitization effects of flavopiridol: Flavopiridol enhanced the cytotoxic effects of paclitaxel, cytarabine, topotecan, doxorubicin, and etoposide only when administered after treatment; only flavopiridol plus cisplatin showed sequence-independent synergy (29). Flavopiridol sensitized retinoblastoma protein-deficient sarcoma cells to doxorubicin-induced cell killing (101) and colon cancer xenografts cells to CPT-11 through activation of caspase 3 and cleavage of anti-apoptotic factors (123). Furthermore, flavopiridol enhanced the induction of apoptosis by gemcitabine in human pancreatic, gastric, and colon cancer cell lines (77). The mechanism accounting for the sensitization effects of flavopiridol may involve its inhibition of NF- $\kappa$ B. Indeed, in one study, flavopiridol inhibited NF- $\kappa$ B, which in turn down-regulated cyclin D1, COX-2, and MMP-9 (187). Similarly, flavopiridol inhibited NF- $\kappa$ B-dependent gene transcription and enhanced TNF- and TNF-related apoptosis-inducing ligand-induced cytotoxicity (85). Yet another study showed that flavopiridol enhanced tumor cell apoptosis by inhibiting survivin phosphorylation (201).

### Silymarin

Silymarin may potentiate the effects of chemotherapy on cancer. In MDR breast cancer cells, it potentiated doxorubicin cytotoxicity by inhibiting P-glycoprotein ATPase activity, which is responsible for cellular efflux of cytotoxic substances (214). This observation was consistent with a recent report demonstrating that silymarin and resveratrol significantly increased breast cancer-resistant protein substrates in breast cancer-resistant protein-overexpressing cells (44). Another study demonstrated the synergistic effects of silybin plus cisplatin or doxorubicin in breast cancer and ovarian cancer cells (164). Likewise, silybin strongly synergized the growth-inhibitory effect of doxorubicin in prostate cancer cells and was associated with cell cycle arrest (195).

## SENSITIZATION OF TUMOR CELLS BY POLYPHENOLS TO RADIATION

### Genistein

Genistein and radiation have been suggested to be synergistic. In prostate cancer cells, genistein significantly inhibited DNA synthesis, cell growth, and colony formation and potentiated the effect of low doses of photon (200–300 cGy) or neutron (100–150 cGy) radiation, the latter effect being more pronounced with the combined treatment than with genistein or radiation alone (65). In an *in vivo* murine lung cancer model, chemoradiotherapy plus genistein resulted in significantly lower tumor volume, microvessel density, and vascular endothelial growth factor level than did



chemoradiotherapy alone (111). In human esophageal cancer cell lines, genistein greatly enhanced radiosensitivity by suppressing radiation-induced activation of the survival signals Akt and p42/p44 extracellular signal-regulated protein kinase (4). Finally, in a Bcr/abl-positive leukemic cell line, genistein increased radiation-induced apoptosis and promoted arrest of the G<sub>2</sub> phase cell cycle (138).

### *Resveratrol*

Resveratrol may induce radiosensitization. Reports have suggested that increased activation of NF- $\kappa$ B and up-regulation of NF- $\kappa$ B-mediated expression of COX-2 and of 5-lipoxygenase (5-LOX) contribute significantly to radioresistance (39, 190) and that inhibition of NF- $\kappa$ B, COX-2, and 5-LOX induces radiosensitization (30, 149, 157). Resveratrol has been shown to down-regulate NF- $\kappa$ B (109), COX-2 (177), and 5-LOX (108). Using clonogenic cell survival assays, one study showed that pretreatment with resveratrol enhanced HeLa and SiHa cell killing and induced an early S phase cell-cycle checkpoint arrest after the cells were exposed to ionizing radiation (216).

### *Curcumin*

Resistance to radiation was suggested in one study to be caused by increased expression of NF- $\kappa$ B-induced prosurvival genes, such as *Bcl-2* and *Bcl-xL*, in response to radiation (124). In that investigation, curcumin plus radiation inhibited NF- $\kappa$ B activation in DU-145 and LNCaP prostate cancer cells and resulted in down-regulation of *Bcl-2*, and curcumin alone enhanced caspase activation and cytochrome *c* release in both cell types, leading to increased apoptosis. Curcumin has also been shown to sensitize PC3 prostate cancer cells to radiation (41).

### *Green tea*

A few studies have suggested that EGCG could modulate radiotherapy. A study using human embryonic endothelial cells found that ionizing radiation induced expression of proangiogenesis genes *membrane type 1-MMP* and *caveolin-1* and that this expression was abrogated by pretreatment with EGCG (9). These results demonstrated that radiotherapy, which is better known for its ability to promote cancer cell death, may increase the expression of NF- $\kappa$ B-mediated target genes that may be inhibited by EGCG (48, 143). Another investigation showed that EGCG suppressed oncogenic transformation of mouse embryonic fibroblast C3H10T1/2 cells (90).

### *Flavopiridol*

Growing evidence suggests that flavopiridol may sensitize tumor cells to radiation. In one study, flavopiridol significantly enhanced the induction of apoptosis by radiation in colon and gastric cancer cells; this effect was optimal when flavopiridol followed the radiation treatment (78). Other investigations showed that flavopiridol strongly enhanced the response of ovarian carcinoma cells to radiation and suggested that the underlying mechanisms included inhibition of

sublethal DNA damage repair and cell cycle redistribution; in addition, transcriptional regulation by flavopiridol may have been involved (78).

## **POLYPHENOLS PROTECTS NORMAL CELLS/ORGANS FROM CHEMO/RADIOTHERAPY**

### *Curcumin*

Several reports have also identified curcumin as a potent protector against radiation. Oral administration of curcumin (200  $\mu$ mol/kg) significantly reduced lung toxicity in rats treated with whole-body radiation (10 Gy in five fractions) (191), and curcumin reduced liver and serum lipid peroxidation in rats treated with radiation. In addition, reduced radiation-induced genotoxicity (chromosomal damage) was observed in mice treated with oral curcumin (400 mmol/kg) plus whole-body radiation (1.5–3.0 Gy) (192). In another study, dietary curcumin significantly decreased acute toxicity from whole-body radiation in rats and decreased the incidence of mammary and pituitary tumors at 1-year follow-up (71). These findings, when taken together, demonstrated that curcumin is a promising radioprotective agent and warrants further investigation.

Curcumin has been shown to protect cells from doxorubicin-induced renal injury (200), cardiotoxicity (199), and gastrointestinal injury (196). These protective effects could be secondary to the suppression of oxidative stress and inflammatory damage (80). One study revealed that curcumin inhibited camptothecin-, mechlorethamine-, and doxorubicin-induced apoptosis in tissue cultures of MCF-7, MDA-MB-231, and BT-474 human breast cancer cells by up to 70%; inhibition occurred after relatively brief 3-h exposures to curcumin, or at curcumin concentrations of 1  $\mu$ M (176). In an *in vivo* model of human breast cancer, dietary supplementation with curcumin significantly inhibited cyclophosphamide-induced tumor regression, and it was concluded that dietary curcumin could inhibit chemotherapy-induced apoptosis by inhibiting reactive oxygen species generation and blocking of c-Jun N-terminal kinase function. However, because NF- $\kappa$ B is a major regulator of the transcriptional response to cellular oxidative stress (54), it is tempting to believe that the suppression of NF- $\kappa$ B-induced gene expression is responsible, at least in part, for these therapeutic effects.

### *CAPE*

Several reports have indicated that CAPE has strong chemoprotective properties. Our investigation demonstrated that pretreatment with CAPE significantly attenuated doxorubicin-induced cardiotoxicity in rats and suggested that these effects may have been due to its antioxidant properties (51). The free-oxygen-radical scavenging properties of CAPE in rats account for its protective effects on renal tissue treated with cisplatin (135) and against lung toxicity after exposure to bleomycin (136). CAPE also protected rat renal cells from ischemia-reperfusion injury (61). CAPE has been shown to

inhibit NF- $\kappa$ B (127), which may account for its chemopreventive effects in intestinal, colon, skin, and liver cancers and reduce inflammation in rats exposed to radiation (106). Furthermore, CAPE suppressed COX-2 expression in human oral epithelial cells and in a rat model of inflammation (115). It may also help to prevent cancer progression by inhibiting angiogenesis, tumor invasion, and metastasis in mice (104) and, when combined with sulindac, by suppressing lung adenocarcinoma motility (171).

### *Silymarin*

Silymarin appears to protect rat cardiomyocytes against anthracycline-induced oxidative stress, perhaps through its effects on cell membrane stabilization (176). These results were consistent with those from another report that suggested silymarin may prevent doxorubicin-mediated damage to rat heart membrane, possibly through a free-radical scavenging mechanism (148).

### *Ginger*

Ginger may prevent chemotherapy and radiotherapy-induced nausea. In shrews, rats, and dogs, ginger inhibited chemotherapy-induced gastric emptying and showed comparable results to standard antiemetic compounds (167, 168, 207). Furthermore, ginger reduced radiation-related sickness and free-radical production in mice (72).

## ENHANCEMENT OF DRUG BIOAVAILABILITY BY POLYPHENOLS

### *Piperine*

Piperine, an ingredient of black pepper, increases the bioavailability of chemotherapeutic drugs. A study using hepatic tissue in rats demonstrated that piperine was a nonspecific inhibitor of drug metabolism and discriminated little between different cytochrome P-450 forms (13), and another study found that it modified the rate of glucuronidation by lowering endogenous UDP-glucuronic acid content and inhibiting transferase activity (174). This observation was consistent with that of another report that suggested piperine was a potent inhibitor of UDP-glucose dehydrogenase in rats and that its effect on glucuronidation was stronger in the intestines than in the liver (154). In a clinical crossover study in which six subjects in each group received a single dose of propranolol or theophylline alone or piperine, piperine enhanced the systemic availability of oral propranolol and theophylline (15). In rat jejunum cells, piperine significantly stimulated  $\gamma$ -glutamyl transpeptidase activity, enhanced the uptake of radiolabeled L-leucine, L-isoleucine, and L-valine, and increased lipid peroxidation, suggesting that piperine may increase intestinal permeability through interaction with the mucosal lipid environment (76). These results were consistent with those from studies that showed that piperine modulates membrane dynamics and permeation characteristics and results in an increase in the small intestine absorptive surface, thereby enabling efficient permeation through the epithelial barrier (82, 83). Piperine was also shown to inhibit

both the drug transporter P-glycoprotein and the major drug-metabolizing enzyme CYP3A4, *in vitro*, indicating that dietary piperine could affect plasma concentrations of P-glycoprotein and CYP3A4 substrates in humans (22). Using rat liver, another study showed that piperine markedly inhibited aflatoxin metabolism *in vitro* and *in vivo*, suggesting enhanced aflatoxin bioavailability (6). Piperine was also shown to enhance curcumin bioavailability in both rats and humans (173). These results, when taken together, warrant the further investigation of piperin as a supplement to chemotherapeutic drugs to enhance their bioavailability.

## CONCLUSIONS

Cancer is a multifactorial disease that, in many cases, requires multimodal therapy, including chemotherapy and radiotherapy. The mechanisms for modulation of signaling pathways that account for the efficacy of these treatments have become increasingly clear in recent years, leading to the clinical and laboratory study of numerous biologic modifiers that may increase their potency and reduce their adverse effects. Despite extensive use for thousands of years because of their alleged medicinal value, plant polyphenols have only recently received significant attention in the literature for their ability to modulate a number of signaling pathways that lead to the initiation and progression of cancer. The ability of these products to sensitize tumor cells to chemotherapy and radiotherapy often correlates with their ability to inhibit well-studied molecular markers. The data are still limited, but the results appear promising, and the use of plant polyphenols as potential modulators of chemotherapy and radiotherapy deserves further investigation.

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## ABBREVIATIONS

CAPE, caffeic acid phenethyl ester; COX-2, cyclooxygenase-2; EGCG, (–)-epigallocatechin gallate; EGF, epidermal growth factor; EGFR, epidermal growth factor receptor; 5-FU, 5-fluorouracil; IAP, inhibitor of apoptosis protein; I $\kappa$ B $\alpha$ , inhibitory subunit of nuclear factor- $\kappa$ B; IL-6, interleukin-6; iNOS, inducible nitric oxide synthase; 5-LOX, 5-lipoxygenase; MDR, multidrug resistance; MMP, matrix metalloproteinase; NF- $\kappa$ B, nuclear factor- $\kappa$ B; PI-3K, phosphatidylinositol 3-kinase; ROI, reactive oxygen intermediates; STAT, signal transducer and activator of transcription; TNF, tumor necrosis factor.

## REFERENCES

- Aggarwal BB, Kumar A, and Bharti AC. Anticancer potential of curcumin: preclinical and clinical studies. *Anticancer Res* 23: 363–398, 2003.
- Ahmad KA, Clement MV, Hanif IM, and Pervaiz S. Resveratrol inhibits drug-induced apoptosis in human leukemia cells by creating an intracellular milieu nonpermissive for death execution. *Cancer Res* 64: 1452–1459, 2004.
- Akimoto T, Hunter NR, Buchmiller L, Mason K, Ang KK, and Milas L. Inverse relationship between epidermal growth factor receptor expression and radiocurability of murine carcinomas. *Clin Cancer Res* 5: 2884–2890, 1999.
- Akimoto T, Nonaka T, Ishikawa H, Sakurai H, Saitoh JI, Takahashi T, and Mitsunashi N. Genistein, a tyrosine kinase inhibitor, enhanced radiosensitivity in human esophageal cancer cell lines in vitro: possible involvement of inhibition of survival signal transduction pathways. *Int J Radiat Oncol Biol Phys* 50: 195–201, 2001.
- Akira S. Roles of STAT3 defined by tissue-specific gene targeting. *Oncogene* 19: 2607–2611, 2000.
- Allameh A, Saxena M, Biswas G, Raj HG, Singh J, and Srivastava N. Piperine, a plant alkaloid of the piper species, enhances the bioavailability of aflatoxin B1 in rat tissues. *Cancer Lett* 61: 195–199, 1992.
- Altieri DC. Validating survivin as a cancer therapeutic target. *Nat Rev Cancer* 3: 46–54, 2003.
- Altieri DC, Marchisio PC, and Marchisio C. Survivin apoptosis: an interloper between cell death and cell proliferation in cancer. *Lab Invest* 79: 1327–1333, 1999.
- Annabi B, Lee YT, Martel C, Pilorget A, Bahary JP, and Beliveau R. Radiation induced-tubulogenesis in endothelial cells is antagonized by the antiangiogenic properties of green tea polyphenol (–) epigallocatechin-3-gallate. *Cancer Biol Ther* 2: 642–649, 2003.
- Anuchapreeda S, Leechanachai P, Smith MM, Ambudkar SV, and Limtrakul PN. Modulation of P-glycoprotein expression and function by curcumin in multidrug-resistant human KB cells. *Biochem Pharmacol* 64: 573–582, 2002.
- Arlt A, Gehrz A, Muerkoster S, Vorndamm J, Kruse ML, Folsch UR, and Schafer H. Role of NF-kappaB and Akt/PI3K in the resistance of pancreatic carcinoma cell lines against gemcitabine-induced cell death. *Oncogene* 22: 3243–3251, 2003.
- Asanuma K, Kobayashi D, Furuya D, Tsuji N, Yagihashi A, and Watanabe N. A role for survivin in radioresistance of pancreatic cancer cells. *Jpn J Cancer Res* 93: 1057–1062, 2002.
- Atal CK, Dubey RK, and Singh J. Biochemical basis of enhanced drug bioavailability by piperine: evidence that piperine is a potent inhibitor of drug metabolism. *J Pharmacol Exp Ther* 232: 258–262, 1985.
- Aziz MH, Kumar R, and Ahmad N. Cancer chemoprevention by resveratrol: in vitro and in vivo studies and the underlying mechanisms. *Int J Oncol* 23: 17–28, 2003.
- Bano G, Raina RK, Zutshi U, Bedi KL, Johri RK, and Sharma SC. Effect of piperine on bioavailability and pharmacokinetics of propranolol and theophylline in healthy volunteers. *Eur J Clin Pharmacol* 41: 615–617, 1991.
- Bao R, Connolly DC, Murphy M, Green J, Weinstein JK, Pisarcik DA, and Hamilton TC. Activation of cancer-specific gene expression by the survivin promoter. *J Natl Cancer Inst* 94: 522–528, 2002.
- Bargou RC, Emmerich F, Krappmann D, Bommert K, Ma-pa MY, Arnold W, Royer HD, Grinstein E, Greiner A, Scheidereit C, and Dorken B. Constitutive nuclear factor-kappaB-RelA activation is required for proliferation and survival of Hodgkin's disease tumor cells. *J Clin Invest* 100: 2961–2969, 1997.
- Barkett M and Gilmore TD. Control of apoptosis by Rel/NF-kappaB transcription factors. *Oncogene* 18: 6910–6924, 1999.
- Baxa DM and Yoshimura FK. Genistein reduces NF-kappa B in T lymphoma cells via a caspase-mediated cleavage of I kappa B alpha. *Biochem Pharmacol* 66: 1009–1018, 2003.
- Beg AA and Baltimore D. An essential role for NF-kappaB in preventing TNF-alpha-induced cell death. *Science* 274: 782–784, 1996.
- Belka C and Budach W. Anti-apoptotic Bcl-2 proteins: structure, function and relevance for radiation biology. *Int J Radiat Biol* 78: 643–658, 2002.
- Bhardwaj RK, Glaeser H, Becquemont L, Klotz U, Gupta SK, and Fromm MF. Piperine, a major constituent of black pepper, inhibits human P-glycoprotein and CYP3A4. *J Pharmacol Exp Ther* 302: 645–650, 2002.
- Bharti AC and Aggarwal BB. Chemopreventive agents induce suppression of nuclear factor-kappaB leading to chemosensitization. *Ann N Y Acad Sci* 973: 392–395, 2002.
- Bharti AC and Aggarwal BB. Nuclear factor-kappa B and cancer: its role in prevention and therapy. *Biochem Pharmacol* 64: 883–888, 2002.
- Bharti AC, Donato N, Singh S, and Aggarwal BB. Curcumin (diferuloylmethane) down-regulates the constitutive activation of nuclear factor-kappa B and IkappaBalpha kinase in human multiple myeloma cells, leading to suppression of proliferation and induction of apoptosis. *Blood* 101: 1053–1062, 2003.
- Bharti AC, Shishodia S, Reuben JM, Weber D, Alexanian R, Raj-Vadhan S, Estrov Z, Talpaz M, and Aggarwal BB. Nuclear factor-kappaB and STAT3 are constitutively active in CD138+ cells derived from multiple myeloma patients, and suppression of these transcription factors leads to apoptosis. *Blood* 103: 3175–3184, 2004.
- Bharti AC, Takada Y, and Aggarwal BB. Curcumin (diferuloylmethane) inhibits receptor activator of NF-kappaB ligand-induced NF-kappaB activation in osteoclast precursors and suppresses osteoclastogenesis. *J Immunol* 172: 5940–5947, 2004.
- Bianco C, Bianco R, Tortora G, Damiano V, Guerrieri P, Montemaggi P, Mendelsohn J, De Placido S, Bianco AR, and Ciardiello F. Antitumor activity of combined treatment of human cancer cells with ionizing radiation and anti-epidermal growth factor receptor monoclonal antibody C225 plus type I protein kinase A antisense oligonucleotide. *Clin Cancer Res* 6: 4343–4350, 2000.
- Bible KC and Kaufmann SH. Cytotoxic synergy between flavopiridol (NSC 649890, L86-8275) and various anti-

- neoplastic agents: the importance of sequence of administration. *Cancer Res* 57: 3375–3380, 1997.
30. Borisova IG and Budnitskaia EV. In vitro radiation sensitivity of lipoxygenase systems. *Radiobiologiya* 21: 58–62, 1981.
  31. Bours V, Dejardin E, Goujon-Letawe F, Merville MP, and Castronovo V. The NF-kappa-B transcription factor and cancer: high expression of NF-kappaB- and IkappaB-related proteins in tumor cell lines. *Biochem Pharmacol* 47: 145–149, 1994.
  32. Bromberg JF. Activation of STAT proteins and growth control. *Bioessays* 23: 161–169, 2001.
  33. Brzozowski AM, Pike AC, Dauter Z, Hubbard RE, Bonn T, Engstrom O, Ohman L, Greene GL, Gustafsson JA, and Carlquist M. Molecular basis of agonism and antagonism in the oestrogen receptor. *Nature* 389: 753–758, 1997.
  34. Buchler P, Gukovskaya AS, Mouria M, Buchler MC, Buchler MW, Friess H, Pandol SJ, Reber HA, and Hines OJ. Prevention of metastatic pancreatic cancer growth in vivo by induction of apoptosis with genistein, a naturally occurring isoflavonoid. *Pancreas* 26: 264–273, 2003.
  35. Cal C, Garban H, Jazirehi A, Yeh C, Mizutani Y, and Bonavida B. Resveratrol and cancer: chemoprevention, apoptosis, and chemo-immunosensitizing activities. *Curr Med Chem Anti-Canc Agents* 3: 77–93, 2003.
  36. Calvin DP, Nam S, Buettner R, Sekharam M, Torres-Roca J, and Jove R. Inhibition of STAT3 activity with STAT3 antisense oligonucleotide (STAT3-ASO) enhances radiation-induced apoptosis in DU145 prostate cancer cells. *Int J Radiat Oncol Biol Phys* 57(Suppl): S297, 2003.
  37. Chandel NS, Schumacker PT, and Arch RH. Reactive oxygen species are downstream products of TRAF-mediated signal transduction. *J Biol Chem* 276: 42728–42736, 2001.
  38. Chen CC, Sun YT, Chen JJ, and Chiu KT. TNF-alpha-induced cyclooxygenase-2 expression in human lung epithelial cells: involvement of the phospholipase C-gamma 2, protein kinase C-alpha, tyrosine kinase, NF-kappa B-inducing kinase, and I-kappa B kinase 1/2 pathway. *J Immunol* 165: 2719–2728, 2000.
  39. Chen X, Shen B, Xia L, Khaletskiy A, Chu D, Wong JY, and Li JJ. Activation of nuclear factor kappaB in radioreistance of TP53-inactive human keratinocytes. *Cancer Res* 62: 1213–1221, 2002.
  40. Chen YC, Shen SC, Lee WR, Hsu FL, Lin HY, Ko CH, and Tseng SW. Emodin induces apoptosis in human promyelocytic HL-60 cells accompanied by activation of caspase 3 cascade but independent of reactive oxygen species production. *Biochem Pharmacol* 64: 1713–1724, 2002.
  41. Chendil D, Ranga RS, Meigooni D, Sathishkumar S, and Ahmed MM. Curcumin confers radiosensitizing effect in prostate cancer cell line PC-3. *Oncogene* 23: 1599–1607, 2004.
  42. Chlopčikova S, Psotova J, Miketova P, and Simanek V. Chemoprotective effect of plant phenolics against anthracycline-induced toxicity on rat cardiomyocytes. Part I. Silymarin and its flavonolignans. *Phytother Res* 18: 107–110, 2004.
  43. Chuang SE, Yeh PY, Lu YS, Lai GM, Liao CM, Gao M, and Cheng AL. Basal levels and patterns of anticancer drug-induced activation of nuclear factor-kappaB (NF-kappaB), and its attenuation by tamoxifen, dexamethasone, and curcumin in carcinoma cells. *Biochem Pharmacol* 63: 1709–1716, 2002.
  44. Cooray HC, Janvilisri T, van Veen HW, Hladky SB, and Barrand MA. Interaction of the breast cancer resistance protein with plant polyphenols. *Biochem Biophys Res Commun* 317: 269–275, 2004.
  45. Cusack JC, Liu R, and Baldwin AS. NF-kappa B and chemoresistance: potentiation of cancer drugs via inhibition of NF-kappa B. *Drug Resist Updat* 2: 271–273, 1999.
  46. Cusack JC Jr, Liu R, and Baldwin AS Jr. Inducible chemoresistance to 7-ethyl-10-[4-(1-piperidino)-1-piperidino]-carbonyloxycamptothecin (CPT-11) in colorectal cancer cells and a xenograft model is overcome by inhibition of nuclear factor-kappaB activation. *Cancer Res* 60: 2323–2330, 2000.
  47. Davis JN, Kucuk O, and Sarkar FH. Genistein inhibits NF-kappa B activation in prostate cancer cells. *Nutr Cancer* 35: 167–174, 1999.
  48. Deregowski V, Delhalle S, Benoit V, Bours V, and Merville MP. Identification of cytokine-induced nuclear factor-kappaB target genes in ovarian and breast cancer cells. *Biochem Pharmacol* 64: 873–881, 2002.
  49. Di Pietro A, Conseil G, Perez-Victoria JM, Dayan G, Baubichon-Cortay H, Trompier D, Steinfels E, Jault JM, de Wet H, Maitrejean M, Comte G, Boumendjel A, Mariotte AM, Dumontet C, McIntosh DB, Goffeau A, Castanys S, Gamarro F, and Barron D. Modulation by flavonoids of cell multidrug resistance mediated by P-glycoprotein and related ABC transporters. *Cell Mol Life Sci* 59: 307–322, 2002.
  50. Dong Z. Molecular mechanism of the chemopreventive effect of resveratrol. *Mutat Res* 523–524: 145–150, 2003.
  51. Fadillioglu E, Oztas E, Erdogan H, Yagmurca M, Sogut S, Ucar M, and Irmak MK. Protective effects of caffeic acid phenethyl ester on doxorubicin-induced cardiotoxicity in rats. *J Appl Toxicol* 24: 47–52, 2004.
  52. Fenig E, Nordenberg J, Beery E, Sulkes J, and Wasserman L. Combined effect of aloe-emodin and chemotherapeutic agents on the proliferation of an adherent variant cell line of Merkel cell carcinoma. *Oncol Rep* 11: 213–217, 2004.
  53. Flynn V Jr, Ramanitharan A, Moparty K, Davis R, Sikka S, Agrawal KC, and Abdel-Mageed AB. Adenovirus-mediated inhibition of NF-kappaB confers chemo-sensitization and apoptosis in prostate cancer cells. *Int J Oncol* 23: 317–323, 2003.
  54. Garg A and Aggarwal BB. Nuclear transcription factor-kappaB as a target for cancer drug development. *Leukemia* 16: 1053–1068, 2002.
  55. Garg AK and Aggarwal BB. Reactive oxygen intermediates in TNF signaling. *Mol Immunol* 39: 509–517, 2002.
  56. Garg AK, Hortobagyi GN, Aggarwal BB, Sahin AA, and Buchholz TA. Nuclear factor-kappaB as a predictor of treatment response in breast cancer. *Curr Opin Oncol* 15: 405–411, 2003.
  57. Gigante M, Toffoli G, Bertola A, Biscontin G, Dassie A, Zanelli GD, Zanin E, Trovo MG, and Muzzio PC. Radiosensitivity in multidrug-resistant and cisplatin-resistant



- human carcinoma cell lines. *Am J Clin Oncol* 26: e73–e79, 2003.
58. Gong L, Li Y, Nedeljkovic-Kurepa A, and Sarkar FH. Inactivation of NF-kappaB by genistein is mediated via Akt signaling pathway in breast cancer cells. *Oncogene* 22: 4702–4709, 2003.
  59. Greten FR, Weber CK, Greten TF, Schneider G, Wagner M, Adler G, and Schmid RM. Stat3 and NF-kappaB activation prevents apoptosis in pancreatic carcinogenesis. *Gastroenterology* 123: 2052–2063, 2002.
  60. Gupta S, Hastak K, Afaq F, Ahmad N, and Mukhtar H. Essential role of caspases in epigallocatechin-3-gallate-mediated inhibition of nuclear factor kappa B and induction of apoptosis. *Oncogene* 23: 2507–2522, 2004.
  61. Gurel A, Armutcu F, Sahin S, Sogut S, Ozyurt H, Gulec M, Kutlu NO, and Akyol O. Protective role of alpha-tocopherol and caffeic acid phenethyl ester on ischemia-reperfusion injury via nitric oxide and myeloperoxidase in rat kidneys. *Clin Chim Acta* 339: 33–41, 2004.
  62. Gutierrez-Puente Y, Zapata-Benavides P, Tari AM, and Lopez-Berestein G. Bcl-2-related antisense therapy. *Semin Oncol* 29: 71–76, 2002.
  63. Hastak K, Gupta S, Ahmad N, Agarwal MK, Agarwal ML, and Mukhtar H. Role of p53 and NF-kappaB in epigallocatechin-3-gallate-induced apoptosis of LNCaP cells. *Oncogene* 22: 4851–4859, 2003.
  64. Hengstler JG, Lange J, Kett A, Dornhofer N, Meinert R, Arand M, Knapstein PG, Becker R, Oesch F, and Tanner B. Contribution of c-erbB-2 and topoisomerase IIalpha to chemoresistance in ovarian cancer. *Cancer Res* 59: 3206–3214, 1999.
  65. Hillman GG, Forman JD, Kucuk O, Yudelev M, Maughan RL, Rubio J, Layer A, Tekyi-Mensah S, Abrams J, and Sarkar FH. Genistein potentiates the radiation effect on prostate carcinoma cells. *Clin Cancer Res* 7: 382–390, 2001.
  66. Hong J, Lambert JD, Lee SH, Sinko PJ, and Yang CS. Involvement of multidrug resistance-associated proteins in regulating cellular levels of (-)-epigallocatechin-3-gallate and its methyl metabolites. *Biochem Biophys Res Commun* 310: 222–227, 2003.
  67. Hour TC, Chen J, Huang CY, Guan JY, Lu SH, and Pu YS. Curcumin enhances cytotoxicity of chemotherapeutic agents in prostate cancer cells by inducing p21(WAF1/CIP1) and C/EBPbeta expressions and suppressing NF-kappaB activation. *Prostate* 51: 211–218, 2002.
  68. Hovelmann S, Beckers TL, and Schmidt M. Molecular alterations in apoptotic pathways after PKB/Akt-mediated chemoresistance in NCI H460 cells. *Br J Cancer* 90: 2370–2377, 2004.
  69. Huang SM and Harari PM. Modulation of radiation response after epidermal growth factor receptor blockade in squamous cell carcinomas: inhibition of damage repair, cell cycle kinetics, and tumor angiogenesis. *Clin Cancer Res* 6: 2166–2174, 2000.
  70. Huang SM, Bock JM, and Harari PM. Epidermal growth factor receptor blockade with C225 modulates proliferation, apoptosis, and radiosensitivity in squamous cell carcinomas of the head and neck. *Cancer Res* 59: 1935–1940, 1999.
  71. Inano H and Onoda M. Radioprotective action of curcumin extracted from *Curcuma longa* Linn: inhibitory effect on formation of urinary 8-hydroxy-2'-deoxyguanosine, tumorigenesis, but not mortality, induced by gamma-ray irradiation. *Int J Radiat Oncol Biol Phys* 53: 735–743, 2002.
  72. Jagetia GC, Baliga MS, Venkatesh P, and Ulloor JN. Influence of ginger rhizome (*Zingiber officinale* Rose) on survival, glutathione and lipid peroxidation in mice after whole-body exposure to gamma radiation. *Radiat Res* 160: 584–592, 2003.
  73. Jang JH and Surh YJ. Protective effect of resveratrol on beta-amyloid-induced oxidative PC12 cell death. *Free Radic Biol Med* 34: 1100–1110, 2003.
  74. Jazirehi AR and Bonavida B. Resveratrol modifies the expression of apoptotic regulatory proteins and sensitizes non-Hodgkin's lymphoma and multiple myeloma cell lines to paclitaxel-induced apoptosis. *Mol Cancer Ther* 3: 71–84, 2004.
  75. Jin W, Wu L, Liang K, Liu B, Lu Y, and Fan Z. Roles of the PI-3K and MEK pathways in Ras-mediated chemoresistance in breast cancer cells. *Br J Cancer* 89: 185–191, 2003.
  76. Johri RK, Thusu N, Khajuria A, and Zutshi U. Piperine-mediated changes in the permeability of rat intestinal epithelial cells. The status of gamma-glutamyl transpeptidase activity, uptake of amino acids and lipid peroxidation. *Biochem Pharmacol* 43: 1401–1407, 1992.
  77. Jung CP, Motwani MV, and Schwartz GK. Flavopiridol increases sensitization to gemcitabine in human gastrointestinal cancer cell lines and correlates with down-regulation of ribonucleotide reductase M2 subunit. *Clin Cancer Res* 7: 2527–2536, 2001.
  78. Jung C, Motwani M, Kortmansky J, Sirotiak FM, She Y, Gonen M, Haimovitz-Friedman A, and Schwartz GK. The cyclin-dependent kinase inhibitor flavopiridol potentiates gamma-irradiation-induced apoptosis in colon and gastric cancer cells. *Clin Cancer Res* 9: 6052–6061, 2003.
  79. Jung M and Dritschilo A. NF-kappa B signaling pathway as a target for human tumor radiosensitization. *Semin Radiat Oncol* 11: 346–351, 2001.
  80. Kapoor S and Priyadarsini KI. Protection of radiation-induced protein damage by curcumin. *Biophys Chem* 92: 119–126, 2001.
  81. Kaufmann SH and Vaux DL. Alterations in the apoptotic machinery and their potential role in anticancer drug resistance. *Oncogene* 22: 7414–7430, 2003.
  82. Khajuria A, Zutshi U, and Bedi KL. Permeability characteristics of piperine on oral absorption—an active alkaloid from peppers and a bioavailability enhancer. *Indian J Exp Biol* 36: 46–50, 1998.
  83. Khajuria A, Thusu N, and Zutshi U. Piperine modulates permeability characteristics of intestine by inducing alterations in membrane dynamics: influence on brush border membrane fluidity, ultrastructure and enzyme kinetics. *Phytomedicine* 9: 224–231, 2002.
  84. Khoshyomn S, Manske GC, Lew SM, Wald SL, and Penar PL. Synergistic action of genistein and cisplatin on growth inhibition and cytotoxicity of human medulloblastoma cells. *Pediatr Neurosurg* 33: 123–131, 2000.
  85. Kim DM, Koo SY, Jeon K, Kim MH, Lee J, Hong CY, and Jeong S. Rapid induction of apoptosis by combination of

- flavopiridol and tumor necrosis factor (TNF)- $\alpha$  or TNF-related apoptosis-inducing ligand in human cancer cell lines. *Cancer Res* 63: 621–626, 2003.
86. Kim R, Tanabe K, Uchida Y, Emi M, and Toge T. Effect of Bcl-2 antisense oligonucleotide on drug-sensitivity in association with apoptosis in undifferentiated thyroid carcinoma. *Int J Mol Med* 11: 799–804, 2003.
  87. Kishi K, Petersen S, Petersen C, Hunter N, Mason K, Masferrer JL, Tofilon PJ, and Milas L. Preferential enhancement of tumor radioresponse by a cyclooxygenase-2 inhibitor. *Cancer Res* 60: 1326–1331, 2000.
  88. Knuefermann C, Lu Y, Liu B, Jin W, Liang K, Wu L, Schmidt M, Mills GB, Mendelsohn J, and Fan Z. HER2/PI-3K/Akt activation leads to a multidrug resistance in human breast adenocarcinoma cells. *Oncogene* 22: 3205–3212, 2003.
  89. Koki AT, Leahy KM, and Masferrer JL. Potential utility of COX-2 inhibitors in chemoprevention and chemotherapy. *Expert Opin Investig Drugs* 8: 1623–1638, 1999.
  90. Komatsu K, Tauchi H, Yano N, Endo S, Matsuura S, and Shoji S. Inhibitory action of (–)-epigallocatechin gallate on radiation-induced mouse oncogenic transformation. *Cancer Lett* 112: 135–139, 1997.
  91. Kubota T, Uemura Y, Kobayashi M, and Taguchi H. Combined effects of resveratrol and paclitaxel on lung cancer cells. *Anticancer Res* 23: 4039–4046, 2003.
  92. Kumi-Diaka J and Townsend J. Toxic potential of dietary genistein isoflavone and beta-lapachone on capacitation and acrosome reaction of epididymal spermatozoa. *J Med Food* 6: 201–208, 2003.
  93. Kurland JF and Meyn RE. Protease inhibitors restore radiation-induced apoptosis to Bcl-2-expressing lymphoma cells. *Int J Cancer* 96: 327–333, 2001.
  94. Lai GH, Zhang Z, and Sirica AE. Celecoxib acts in a cyclooxygenase-2-independent manner and in synergy with emodin to suppress rat cholangiocarcinoma growth in vitro through a mechanism involving enhanced Akt inactivation and increased activation of caspases-9 and -3. *Mol Cancer Ther* 2: 265–271, 2003.
  95. Lei W, Mayotte JE, and Levitt ML. Enhancement of chemosensitivity and programmed cell death by tyrosine kinase inhibitors correlates with EGFR expression in non-small cell lung cancer cells. *Anticancer Res* 19: 221–228, 1999.
  96. Leonard GD, Polgar O, and Bates SE. ABC transporters and inhibitors: new targets, new agents. *Curr Opin Investig Drugs* 3: 1652–1659, 2002.
  97. Levy DE and Darnell JE Jr. Stats: transcriptional control and biological impact. *Nat Rev Mol Cell Biol* 3: 651–662, 2002.
  98. Levy DE and Lee CK. What does Stat3 do? *J Clin Invest* 109: 1143–1148, 2002.
  99. Li W and Weber G. Synergistic action of tiazofurin and genistein in human ovarian carcinoma cells. *Oncol Res* 10: 117–122, 1998.
  100. Li W and Weber G. Synergistic action of tiazofurin and genistein on growth inhibition and differentiation of K-562 human leukemic cells. *Life Sci* 63: 1975–1981, 1998.
  101. Li W, Fan J, and Bertino JR. Selective sensitization of retinoblastoma protein-deficient sarcoma cells to doxorubicin by flavopiridol-mediated inhibition of cyclin-dependent kinase 2 kinase activity. *Cancer Res* 61: 2579–2582, 2001.
  102. Li Y and Sarkar FH. Inhibition of nuclear factor kappaB activation in PC3 cells by genistein is mediated via Akt signaling pathway. *Clin Cancer Res* 8: 2369–2377, 2002.
  103. Li Y, Ellis KL, Ali S, El-Rayes BF, Nedeljkovic-Kurepa A, Kucuk O, Philip PA, and Sarkar FH. Apoptosis-inducing effect of chemotherapeutic agents is potentiated by soy isoflavone genistein, a natural inhibitor of NF-kappaB in BxPC-3 pancreatic cancer cell line. *Pancreas* 28: e90–e95, 2004.
  104. Liao HF, Chen YY, Liu JJ, Hsu ML, Shieh HJ, Liao HJ, Shieh CJ, Shiao MS, and Chen YJ. Inhibitory effect of caffeic acid phenethyl ester on angiogenesis, tumor invasion, and metastasis. *J Agric Food Chem* 51: 7907–7912, 2003.
  105. Limtrakul P, Anuchapreeda S, and Buddhasukh D. Modulation of human multidrug-resistance MDR-1 gene by natural curcuminoids. *BMC Cancer* 4: 13, 2004.
  106. Linard C, Marquette C, Mathieu J, Pennequin A, Clarencon D, and Mathe D. Acute induction of inflammatory cytokine expression after gamma-irradiation in the rat: effect of an NF-kappaB inhibitor. *Int J Radiat Oncol Biol Phys* 58: 427–434, 2004.
  107. Lu B, Mu Y, Cao C, Zeng F, Schneider S, Tan J, Price J, Chen J, Freeman M, and Hallahan DE. Survivin as a therapeutic target for radiation sensitization in lung cancer. *Cancer Res* 64: 2840–2845, 2004.
  108. MacCarrone M, Lorenzon T, Guerrieri P, and Agro AF. Resveratrol prevents apoptosis in K562 cells by inhibiting lipoyxygenase and cyclooxygenase activity. *Eur J Biochem* 265: 27–34, 1999.
  109. Manna SK, Mukhopadhyay A, and Aggarwal BB. Resveratrol suppresses TNF-induced activation of nuclear transcription factors NF-kappa B, activator protein-1, and apoptosis: potential role of reactive oxygen intermediates and lipid peroxidation. *J Immunol* 164: 6509–6519, 2000.
  110. Masuda M, Suzui M, Lim JT, Deguchi A, Soh JW, and Weinstein IB. Epigallocatechin-3-gallate decreases VEGF production in head and neck and breast carcinoma cells by inhibiting EGFR-related pathways of signal transduction. *J Exp Ther Oncol* 2: 350–359, 2002.
  111. McDonnell CO, Holden G, Sheridan ME, Foley D, Moriarty M, Walsh TN, and Bouchier-Hayes DJ. Improvement in efficacy of chemoradiotherapy by addition of an antiangiogenic agent in a murine tumor model. *J Surg Res* 116: 19–23, 2004.
  112. McKenna WG and Muschel RJ. Targeting tumor cells by enhancing radiation sensitivity. *Genes Chromosomes Cancer* 38: 330–338, 2003.
  113. Mendelsohn J. Epidermal growth factor receptor inhibition by a monoclonal antibody as anticancer therapy. *Clin Cancer Res* 3: 2703–2707, 1997.
  114. Mesri M, Morales-Ruiz M, Ackermann EJ, Bennett CF, Pober JS, Sessa WC, and Altieri DC. Suppression of vascular endothelial growth factor-mediated endothelial cell protection by survivin targeting. *Am J Pathol* 158: 1757–1765, 2001.

115. Michaluart P, Masferrer JL, Carothers AM, Subbaramaiah K, Zweifel BS, Koboldt C, Mestre JR, Grunberger D, Sacks PG, Tanabe T, and Dannenberg AJ. Inhibitory effects of caffeic acid phenethyl ester on the activity and expression of cyclooxygenase-2 in human oral epithelial cells and in a rat model of inflammation. *Cancer Res* 59: 2347–2352, 1999.
116. Milas L. Cyclooxygenase-2 (COX-2) enzyme inhibitors as potential enhancers of tumor radioresponse. *Semin Radiat Oncol* 11: 290–299, 2001.
117. Milas L and Hanson WR. Eicosanoids and radiation. *Eur J Cancer* 31A: 1580–1585, 1995.
118. Milas L, Kishi K, Hunter N, Mason K, Masferrer JL, and Tofilon PJ. Enhancement of tumor response to gamma-radiation by an inhibitor of cyclooxygenase-2 enzyme. *J Natl Cancer Inst* 91: 1501–1504, 1999.
119. Milas L, Mason K, Hunter N, Petersen S, Yamakawa M, Ang K, Mendelsohn J, and Fan Z. *In vivo* enhancement of tumor radioresponse by C225 antiepidermal growth factor receptor antibody. *Clin Cancer Res* 6: 701–708, 2000.
120. Milas L, Mason KA, Liao Z, and Ang KK. Chemoradiotherapy: emerging treatment improvement strategies. *Head Neck* 25: 152–167, 2003.
121. Mohandas KM and Desai DC. Epidemiology of digestive tract cancers in India. V. Large and small bowel. *Indian J Gastroenterol* 18: 118–121, 1999.
122. Mori N, Fujii M, Ikeda S, Yamada Y, Tomonaga M, Ballard DW, and Yamamoto N. Constitutive activation of NF-kappaB in primary adult T-cell leukemia cells. *Blood* 93: 2360–2368, 1999.
123. Motwani M, Jung C, Sirotinak FM, She Y, Shah MA, Gonen M, and Schwartz GK. Augmentation of apoptosis and tumor regression by flavopiridol in the presence of CPT-11 in Hct116 colon cancer monolayers and xenografts. *Clin Cancer Res* 7: 4209–4219, 2001.
124. Mukhopadhyay A, Bueso-Ramos C, Chatterjee D, Pantazis P, and Aggarwal BB. Curcumin downregulates cell survival mechanisms in human prostate cancer cell lines. *Oncogene* 20: 7597–7609, 2001.
125. Nakata E, Mason KA, Hunter N, Husain A, Raju U, Liao Z, Ang KK, and Milas L. Potentiation of tumor response to radiation or chemoradiation by selective cyclooxygenase-2 enzyme inhibitors. *Int J Radiat Oncol Biol Phys* 58: 369–375, 2004.
126. Nasu S, Ang KK, Fan Z, and Milas L. C225 antiepidermal growth factor receptor antibody enhances tumor radiocurability. *Int J Radiat Oncol Biol Phys* 51: 474–477, 2001.
127. Natarajan K, Singh S, Burke TR Jr, Grunberger D, and Aggarwal BB. Caffeic acid phenethyl ester is a potent and specific inhibitor of activation of nuclear transcription factor NF-kappa B. *Proc Natl Acad Sci U S A* 93: 9090–9095, 1996.
128. Natarajan K, Manna SK, Chaturvedi MM, and Aggarwal BB. Protein tyrosine kinase inhibitors block tumor necrosis factor-induced activation of nuclear factor-kappaB, degradation of IkappaBalpha, nuclear translocation of p65, and subsequent gene expression. *Arch Biochem Biophys* 352: 59–70, 1998.
129. Newton TR, Patel NM, Bhat-Nakshatri P, Stauss CR, Goulet RJ, Jr., and Nakshatri H. Negative regulation of transactivation function but not DNA binding of NF-kappaB and AP-1 by IkappaBbeta1 in breast cancer cells. *J Biol Chem* 274: 18827–18835, 1999.
130. Nicolini G, Rigolio R, Miloso M, Bertelli AA, and Tredici G. Anti-apoptotic effect of trans-resveratrol on paclitaxel-induced apoptosis in the human neuroblastoma SH-SY5Y cell line. *Neurosci Lett* 302: 41–44, 2001.
131. Nomura M, Ma W, Chen N, Bode AM, and Dong Z. Inhibition of 12-O-tetradecanoylphorbol-13-acetate-induced NF-kappaB activation by tea polyphenols, (–)-epigallocatechin gallate and theaflavins. *Carcinogenesis* 21: 1885–1890, 2000.
132. O'Connor DS, Schechner JS, Adida C, Mesri M, Rothermel AL, Li F, Nath AK, Pober JS, and Altieri DC. Control of apoptosis during angiogenesis by survivin expression in endothelial cells. *Am J Pathol* 156: 393–398, 2000.
133. O'Connor DS, Wall NR, Porter AC, and Altieri DC. A p34(cdc2) survival checkpoint in cancer. *Cancer Cell* 2: 43–54, 2002.
134. Okabe S, Fujimoto N, Sueoka N, Suganuma M, and Fujiki H. Modulation of gene expression by (–)-epigallocatechin gallate in PC-9 cells using a cDNA expression array. *Biol Pharm Bull* 24: 883–886, 2001.
135. Ozen S, Akyol O, Iraz M, Sogut S, Ozugurlu F, Ozyurt H, Odaci E, and Yildirim Z. Role of caffeic acid phenethyl ester, an active component of propolis, against cisplatin-induced nephrotoxicity in rats. *J Appl Toxicol* 24: 27–35, 2004.
136. Ozyurt H, Sogut S, Yildirim Z, Kart L, Iraz M, Armutcu F, Temel I, Ozen S, Uzun A, and Akyol O. Inhibitory effect of caffeic acid phenethyl ester on bleomycin-induced lung fibrosis in rats. *Clin Chim Acta* 339: 65–75, 2004.
137. Pahl HL. Activators and target genes of Rel/NF-kappaB transcription factors. *Oncogene* 18: 6853–6866, 1999.
138. Papazisis KT, Zambouli D, Kimoundri OT, Papadakis ES, Vala V, Geromichalos GD, Voyatzis S, Markala D, Destouni E, Boutis L, and Kortsaris AH. Protein tyrosine kinase inhibitor, genistein, enhances apoptosis and cell cycle arrest in K562 cells treated with gamma-irradiation. *Cancer Lett* 160: 107–113, 2000.
139. Park JH, Oh EJ, Choi YH, Kang CD, Kang HS, Kim DK, Kang KI, and Yoo MA. Synergistic effects of dexamethasone and genistein on the expression of Cdk inhibitor p21WAF1/CIP1 in human hepatocellular and colorectal carcinoma cells. *Int J Oncol* 18: 997–1002, 2001.
140. Park OJ and Surh YJ. Chemopreventive potential of epigallocatechin gallate and genistein: evidence from epidemiological and laboratory studies. *Toxicol Lett* 150: 43–56, 2004.
141. Pennati M, Binda M, Colella G, Folini M, Citti L, Villa R, Daidone MG, and Zaffaroni N. Radiosensitization of human melanoma cells by ribozyme-mediated inhibition of survivin expression. *J Invest Dermatol* 120: 648–654, 2003.
142. Petersen C, Petersen S, Milas L, Lang FF, and Tofilon PJ. Enhancement of intrinsic tumor cell radiosensitivity induced by a selective cyclooxygenase-2 inhibitor. *Clin Cancer Res* 6: 2513–2520, 2000.
143. Philip S, Bulbule A, and Kundu GC. Osteopontin stimulates tumor growth and activation of promatrix metallo-

- proteinase-2 through nuclear factor-kappa B-mediated induction of membrane type 1 matrix metalloproteinase in murine melanoma cells. *J Biol Chem* 276: 44926–44935, 2001.
144. Pianetti S, Arsura M, Romieu-Mourez R, Coffey RJ, and Sonenshein GE. Her-2/neu overexpression induces NF-kappaB via a PI3-kinase/Akt pathway involving calpain-mediated degradation of IkappaB-alpha that can be inhibited by the tumor suppressor PTEN. *Oncogene* 20: 1287–1299, 2001.
145. Pianetti S, Guo S, Kavanagh KT, and Sonenshein GE. Green tea polyphenol epigallocatechin-3 gallate inhibits Her-2/neu signaling, proliferation, and transformed phenotype of breast cancer cells. *Cancer Res* 62: 652–655, 2002.
146. Pisters KM, Newman RA, Coldman B, Shin DM, Khuri FR, Hong WK, Glisson BS, and Lee JS. Phase I trial of oral green tea extract in adult patients with solid tumors. *J Clin Oncol* 19: 1830–1838, 2001.
147. Prasad KN. Antioxidants in cancer care: when and how to use them as an adjunct to standard and experimental therapies. *Expert Rev Anticancer Ther* 3: 903–915, 2003.
148. Psotova J, Chlopickova S, Grambal F, Simanek V, and Ulrichova J. Influence of silymarin and its flavonolignans on doxorubicin-iron induced lipid peroxidation in rat heart microsomes and mitochondria in comparison with quercetin. *Phytother Res* 16(Suppl 1): S63–S67, 2002.
149. Pyo H, Choy H, Amorino GP, Kim JS, Cao Q, Hercules SK, and DuBois RN. A selective cyclooxygenase-2 inhibitor, NS-398, enhances the effect of radiation in vitro and in vivo preferentially on the cells that express cyclooxygenase-2. *Clin Cancer Res* 7: 2998–3005, 2001.
150. Raju U, Nakata E, Mason KA, Ang KK, and Milas L. Flavopiridol, a cyclin-dependent kinase inhibitor, enhances radiosensitivity of ovarian carcinoma cells. *Cancer Res* 63: 3263–3267, 2003.
151. Rath PC and Aggarwal BB. Antiproliferative effects of IFN-alpha correlate with the downregulation of nuclear factor-kappa B in human Burkitt lymphoma Daudi cells. *J Interferon Cytokine Res* 21: 523–528, 2001.
152. Rayet B and Gelinas C. Aberrant rel/nfkb genes and activity in human cancer. *Oncogene* 18: 6938–6947, 1999.
153. Real PJ, Sierra A, De Juan A, Segovia JC, Lopez-Vega JM, and Fernandez-Luna JL. Resistance to chemotherapy via Stat3-dependent overexpression of Bcl-2 in metastatic breast cancer cells. *Oncogene* 21: 7611–7618, 2002.
154. Reen RK, Jamwal DS, Taneja SC, Koul JL, Dubey RK, Wiebel FJ, and Singh J. Impairment of UDP-glucose dehydrogenase and glucuronidation activities in liver and small intestine of rat and guinea pig in vitro by piperine. *Biochem Pharmacol* 46: 229–238, 1993.
155. Rosenberg B. Fundamental studies with cisplatin. *Cancer* 55: 2303–2316, 1985.
156. Rosser CJ, Tanaka M, Pisters LL, Tanaka N, Levy LB, Hoover DC, Grossman HB, McDonnell TJ, Kuban DA, and Meyn RE. Adenoviral-mediated PTEN transgene expression sensitizes Bcl-2-expressing prostate cancer cells to radiation. *Cancer Gene Ther* 11: 273–279, 2004.
157. Russo SM, Tepper JE, Baldwin AS Jr, Liu R, Adams J, Elliott P, and Cusack JC Jr. Enhancement of radiosensitivity by proteasome inhibition: implications for a role of NF-kappaB. *Int J Radiat Oncol Biol Phys* 50: 183–193, 2001.
158. Sadzuka Y, Sugiyama T, Miyagishima A, Nozawa Y, and Hirota S. The effects of theanine, as a novel biochemical modulator, on the antitumor activity of adriamycin. *Cancer Lett* 105: 203–209, 1996.
159. Sadzuka Y, Sugiyama T, and Hirota S. Modulation of cancer chemotherapy by green tea. *Clin Cancer Res* 4: 153–156, 1998.
160. Sadzuka Y, Sugiyama T, and Sonobe T. Efficacies of tea components on doxorubicin induced antitumor activity and reversal of multidrug resistance. *Toxicol Lett* 114: 155–162, 2000.
161. Sadzuka Y, Sugiyama T, and Sonobe T. Improvement of idarubicin induced antitumor activity and bone marrow suppression by theanine, a component of tea. *Cancer Lett* 158: 119–124, 2000.
162. Salvesen GS and Duckett CS. IAP proteins: blocking the road to death's door. *Nat Rev Mol Cell Biol* 3: 401–410, 2002.
163. Sarkar FH and Li Y. Mechanisms of cancer chemoprevention by soy isoflavone genistein. *Cancer Metastasis Rev* 21: 265–280, 2002.
164. Scambia G, De Vincenzo R, Ranelletti FO, Panici PB, Ferrandina G, D'Agostino G, Fattorossi A, Bombardelli E, and Mancuso S. Antiproliferative effect of silybin on gynaecological malignancies: synergism with cisplatin and doxorubicin. *Eur J Cancer* 32A: 877–882, 1996.
165. Schwartz GK, Farsi K, Maslak P, Kelsen DP, and Spriggs D. Potentiation of apoptosis by flavopiridol in mitomycin-C-treated gastric and breast cancer cells. *Clin Cancer Res* 3: 1467–1472, 1997.
166. Shabbits JA, Krishna R, and Mayer LD. Molecular and pharmacological strategies to overcome multidrug resistance. *Expert Rev Anticancer Ther* 1: 585–594, 2001.
167. Sharma SS and Gupta YK. Reversal of cisplatin-induced delay in gastric emptying in rats by ginger (Zingiber officinale). *J Ethnopharmacol* 62: 49–55, 1998.
168. Sharma SS, Kochupillai V, Gupta SK, Seth SD, and Gupta YK. Antiemetic efficacy of ginger (Zingiber officinale) against cisplatin-induced emesis in dogs. *J Ethnopharmacol* 57: 93–96, 1997.
169. Shen F and Weber G. Synergistic action of quercetin and genistein in human ovarian carcinoma cells. *Oncol Res* 9: 597–602, 1997.
170. Shi Y. Mechanisms of caspase activation and inhibition during apoptosis. *Mol Cell* 9: 459–470, 2002.
171. Shigeoka Y, Igishi T, Matsumoto S, Nakanishi H, Kodani M, Yasuda K, Hitsuda Y, and Shimizu E. Sulindac sulfide and caffeic acid phenethyl ester suppress the motility of lung adenocarcinoma cells promoted by transforming growth factor-beta through Akt inhibition. *J Cancer Res Clin Oncol* 130: 146–152, 2004.
172. Shishodia S, Majumdar S, Banerjee S, and Aggarwal BB. Ursolic acid inhibits nuclear factor-kappaB activation induced by carcinogenic agents through suppression of IkappaBalpha kinase and p65 phosphorylation: correlation with down-regulation of cyclooxygenase 2, matrix metalloproteinase 9, and cyclin D1. *Cancer Res* 63: 4375–4383, 2003.



173. Shoba G, Joy D, Joseph T, Majeed M, Rajendran R, and Srinivas PS. Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. *Planta Med* 64: 353–356, 1998.
174. Singh J, Dubey RK, and Atal CK. Piperine-mediated inhibition of glucuronidation activity in isolated epithelial cells of the guinea-pig small intestine: evidence that piperine lowers the endogenous UDP-glucuronic acid content. *J Pharmacol Exp Ther* 236: 488–493, 1986.
175. Slichenmyer WJ, Rowinsky EK, Donehower RC, and Kaufmann SH. The current status of camptothecin analogues as antitumor agents. *J Natl Cancer Inst* 85: 271–291, 1993.
176. Somasundaram S, Edmund NA, Moore DT, Small GW, Shi YY, and Orlowski RZ. Dietary curcumin inhibits chemotherapy-induced apoptosis in models of human breast cancer. *Cancer Res* 62: 3868–3875, 2002.
177. Subbaramaiah K, Chung WJ, Michaluart P, Telang N, Tanabe T, Inoue H, Jang M, Pezzuto JM, and Dannenberg AJ. Resveratrol inhibits cyclooxygenase-2 transcription and activity in phorbol ester-treated human mammary epithelial cells. *J Biol Chem* 273: 21875–21882, 1998.
178. Sugiyama T and Sadzuka Y. Combination of theanine with doxorubicin inhibits hepatic metastasis of M5076 ovarian sarcoma. *Clin Cancer Res* 5: 413–416, 1999.
179. Sugiyama T and Sadzuka Y. Enhancing effects of green tea components on the antitumor activity of adriamycin against M5076 ovarian sarcoma. *Cancer Lett* 133: 19–26, 1998.
180. Sugiyama T and Sadzuka Y. Theanine and glutamate transporter inhibitors enhance the antitumor efficacy of chemotherapeutic agents. *Biochim Biophys Acta* 1653: 47–59, 2003.
181. Sugiyama T, Sadzuka Y, Nagasawa K, Ohnishi N, Yokoyama T, and Sonobe T. Membrane transport and antitumor activity of pirarubicin, and comparison with those of doxorubicin. *Jpn J Cancer Res* 90: 775–780, 1999.
182. Surh Y-J. Cancer chemoprevention with dietary phytochemicals. *Nat Rev Cancer* 3: 768–780, 2003.
183. Suzuki Y, Tsubono Y, Nakaya N, Koizumi Y, and Tsuji I. Green tea and the risk of breast cancer: pooled analysis of two prospective studies in Japan. *Br J Cancer* 90: 1361–1363, 2004.
184. Tacchini L, Dansi P, Matteucci E, and Desiderio MA. Hepatocyte growth factor signal coupling to various transcription factors depends on triggering of Met receptor and protein kinase transducers in human hepatoma cells HepG2. *Exp Cell Res* 256: 272–281, 2000.
185. Tainton KM, Smyth MJ, Jackson JT, Tanner JE, Cerruti L, Jane SM, Darcy PK, and Johnstone RW. Mutational analysis of P-glycoprotein: suppression of caspase activation in the absence of ATP-dependent drug efflux. *Cell Death Differ* 11: 1028–1037, 2004.
186. Takada Y and Aggarwal BB. Betulinic acid suppresses carcinogen-induced NF-kappa B activation through inhibition of I kappa B alpha kinase and p65 phosphorylation: abrogation of cyclooxygenase-2 and matrix metalloproteinase-9. *J Immunol* 171: 3278–3286, 2003.
187. Takada Y and Aggarwal BB. Flavopiridol inhibits NF-kappaB activation induced by various carcinogens and inflammatory agents through inhibition of IkappaBalpha kinase and p65 phosphorylation: abrogation of cyclin D1, cyclooxygenase-2, and matrix metalloproteinase-9. *J Biol Chem* 279: 4750–4759, 2004.
188. Tamura S, Bito T, Ichihashi M, and Ueda M. Genistein enhances the cisplatin-induced inhibition of cell growth and apoptosis in human malignant melanoma cells. *Pigment Cell Res* 16: 470–476, 2003.
189. Tanno S, Yanagawa N, Habiro A, Koizumi K, Nakano Y, Osanai M, Mizukami Y, Okumura T, Testa JR, and Kohgo Y. Serine/threonine kinase AKT is frequently activated in human bile duct cancer and is associated with increased radioresistance. *Cancer Res* 64: 3486–3490, 2004.
190. Terakado N, Shintani S, Yano J, Chunnan L, Mihara M, Nakashiro K, and Hamakawa H. Overexpression of cyclooxygenase-2 is associated with radioresistance in oral squamous cell carcinoma. *Oral Oncol* 40: 383–389, 2004.
191. Thresiamma KC, George J, and Kuttan R. Protective effect of curcumin, ellagic acid and bixin on radiation induced toxicity. *Indian J Exp Biol* 34: 845–847, 1996.
192. Thresiamma KC, George J, and Kuttan R. Protective effect of curcumin, ellagic acid and bixin on radiation induced genotoxicity. *J Exp Clin Cancer Res* 17: 431–434, 1998.
193. Tran J, Master Z, Yu JL, Rak J, Dumont DJ, and Kerbel RS. A role for survivin in chemoresistance of endothelial cells mediated by VEGF. *Proc Natl Acad Sci U S A* 99: 4349–4354, 2002.
194. Tsuruo T, Iida H, Tsukagoshi S, and Sakurai Y. 4'-O-Tetrahydropyranyladriamycin as a potential new antitumor agent. *Cancer Res* 42: 1462–1467, 1982.
195. Tyagi AK, Singh RP, Agarwal C, Chan DC, and Agarwal R. Silibinin strongly synergizes human prostate carcinoma DU145 cells to doxorubicin-induced growth inhibition, G2-M arrest, and apoptosis. *Clin Cancer Res* 8: 3512–3519, 2002.
196. Ukil A, Maity S, Karmakar S, Datta N, Vedasiromoni JR, and Das PK. Curcumin, the major component of food flavour turmeric, reduces mucosal injury in trinitrobenzene sulphonic acid-induced colitis. *Br J Pharmacol* 139: 209–218, 2003.
197. van Brussel JP and Mickisch GH. Multidrug resistance in prostate cancer. *Onkologie* 26: 175–181, 2003.
198. Vayalil PK and Katiyar SK. Treatment of epigallocatechin-3-gallate inhibits matrix metalloproteinases-2 and -9 via inhibition of activation of mitogen-activated protein kinases, c-jun and NF-kappaB in human prostate carcinoma DU-145 cells. *Prostate* 59: 33–42, 2004.
199. Venkatesan N. Curcumin attenuation of acute adriamycin myocardial toxicity in rats. *Br J Pharmacol* 124: 425–427, 1998.
200. Venkatesan N, Punithavathi D, and Arumugam V. Curcumin prevents adriamycin nephrotoxicity in rats. *Br J Pharmacol* 129: 231–234, 2000.
201. Wall NR, O'Connor DS, Plescia J, Pommier Y, and Altieri DC. Suppression of survivin phosphorylation on Thr34 by flavopiridol enhances tumor cell apoptosis. *Cancer Res* 63: 230–235, 2003.
202. Wang CY, Mayo MW, and Baldwin AS Jr. TNF- and cancer therapy-induced apoptosis: potentiation by inhibition of NF-kappaB. *Science* 274: 784–787, 1996.

203. Wang CY, Cusack JC Jr, Liu R, and Baldwin AS Jr. Control of inducible chemoresistance: enhanced anti-tumor therapy through increased apoptosis by inhibition of NF-kappaB. *Nat Med* 5: 412–417, 1999.
204. Weaver KD, Yeyeodu S, Cusack JC Jr, Baldwin AS Jr, and Ewend MG. Potentiation of chemotherapeutic agents following antagonism of nuclear factor kappa B in human gliomas. *J Neurooncol* 61: 187–196, 2003.
205. Wietrzyk J, Boratynski J, Grynkiewicz G, Ryczynski A, Radzikowski C, and Opolski A. Antiangiogenic and anti-tumour effects in vivo of genistein applied alone or combined with cyclophosphamide. *Anticancer Res* 21: 3893–3896, 2001.
206. Yamagishi N, Miyakoshi J, and Takebe H. Enhanced radiosensitivity by inhibition of nuclear factor kappa B activation in human malignant glioma cells. *Int J Radiat Biol* 72: 157–162, 1997.
207. Yamahara J, Rong HQ, Naitoh Y, Kitani T, and Fujimura H. Inhibition of cytotoxic drug-induced vomiting in sun-cus by a ginger constituent. *J Ethnopharmacol* 27: 353–355, 1989.
208. Yi J, Yang J, He R, Gao F, Sang H, Tang X, and Ye RD. Emodin enhances arsenic trioxide-induced apoptosis via generation of reactive oxygen species and inhibition of survival signaling. *Cancer Res* 64: 108–116, 2004.
209. Zhan M and Han ZC. Phosphatidylinositol 3-kinase/AKT in radiation responses. *Histol Histopathol* 19: 915–923, 2004.
210. Zhang L and Hung MC. Sensitization of HER-2/neu-overexpressing non-small cell lung cancer cells to chemotherapeutic drugs by tyrosine kinase inhibitor emodin. *Oncogene* 12: 571–576, 1996.
211. Zhang L, Chang CJ, Bacus SS, and Hung MC. Suppressed transformation and induced differentiation of HER-2/neu-overexpressing breast cancer cells by emodin. *Cancer Res* 55: 3890–3896, 1995.
212. Zhang L, Lau YK, Xi L, Hong RL, Kim DS, Chen CF, Hortobagyi GN, Chang C, and Hung MC. Tyrosine kinase inhibitors, emodin and its derivative repress HER-2/neu-induced cellular transformation and metastasis-associated properties. *Oncogene* 16: 2855–2863, 1998.
213. Zhang L, Lau YK, Xia W, Hortobagyi GN, and Hung MC. Tyrosine kinase inhibitor emodin suppresses growth of HER-2/neu-overexpressing breast cancer cells in athymic mice and sensitizes these cells to the inhibitory effect of paclitaxel. *Clin Cancer Res* 5: 343–353, 1999.
214. Zhang S and Morris ME. Effects of the flavonoids biochanin A, morin, phloretin, and silymarin on P-glycoprotein-mediated transport. *J Pharmacol Exp Ther* 304: 1258–1267, 2003.
215. Zhu L, Fukuda S, Cordis G, Das DK, and Maulik N. Anti-apoptotic protein survivin plays a significant role in tubular morphogenesis of human coronary arteriolar endothelial cells by hypoxic preconditioning. *FEBS Lett* 508: 369–374, 2001.
216. Zoberi I, Bradbury CM, Curry HA, Bisht KS, Goswami PC, Roti Roti JL, and Gius D. Radiosensitizing and anti-proliferative effects of resveratrol in two human cervical tumor cell lines. *Cancer Lett* 175: 165–173, 2002.
217. Zwelling LA, Michaels S, Schwartz H, Dobson PP, and Kohn KW. DNA cross-linking as an indicator of sensitivity and resistance of mouse L1210 leukemia to cis-diaminedichloroplatinum(II) and L-phenylalanine mustard. *Cancer Res* 41: 640–649, 1981.

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2. Shu-Chun Hsu, Jing-Gung Chung. 2012. Anticancer potential of emodin. *BioMedicine* **2**:3, 108-116. [[CrossRef](#)]
3. Wei Liu, Qian Feng, Ye Li, Ling Ye, Ming Hu, Zhongqiu Liu. 2012. Coupling of UDP-glucuronosyltransferases and multidrug resistance-associated proteins is responsible for the intestinal disposition and poor bioavailability of emodin. *Toxicology and Applied Pharmacology* . [[CrossRef](#)]
4. Xi Lin, Gang Wu, Wen-Qian Huo, Yao Zhang, Feng-Shuo Jin. 2012. Resveratrol induces apoptosis associated with mitochondrial dysfunction in bladder carcinoma cells. *International Journal of Urology* **19**:8, 757-764. [[CrossRef](#)]
5. FJ Gómez-García, MP López-Jornet, N Álvarez-Sánchez, J Castillo-Sánchez, O Benavente-García, V Vicente Ortega. 2012. Effect of the phenolic compounds apigenin and carnosic acid on oral carcinogenesis in hamster induced by DMBA. *Oral Diseases* n/a-n/a. [[CrossRef](#)]
6. Subash C. Gupta , David Hevia , Sridevi Patchva , Byoungduck Park , Wonil Koh , Bharat B. Aggarwal . 2012. Upsides and Downsides of Reactive Oxygen Species for Cancer: The Roles of Reactive Oxygen Species in Tumorigenesis, Prevention, and Therapy. *Antioxidants & Redox Signaling* **16**:11, 1295-1322. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
7. Zheng-Yun Zou , Jia Wei , Xiao-Lin Li , Li-Xia Yu , Ting-Ting Wang , Xiao-Ping Qian , Bao-Rui Liu . 2012. Enhancement of Anticancer Efficacy of Chemotherapeutics by Gambogic Acid Against Gastric Cancer Cells. *Cancer Biotherapy & Radiopharmaceuticals* **27**:5, 299-306. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
8. NM Rogers, MD Stephenson, AR Kitching, JD Horowitz, PTH Coates. 2012. Amelioration of renal ischaemia-reperfusion injury by liposomal delivery of curcumin to renal tubular epithelial and antigen-presenting cells. *British Journal of Pharmacology* **166**:1, 194-209. [[CrossRef](#)]
9. Soon Young Shin, Ji Ho Kim, Jung Ho Lee, Yoongho Lim, Young Han Lee. 2012. 2#-Hydroxyflavanone induces apoptosis through Egr-1 involving expression of Bax, p21, and NAG-1 in colon cancer cells. *Molecular Nutrition & Food Research* **56**:5, 761-774. [[CrossRef](#)]
10. Yujiang Fang, Vincent G. DeMarco, Michael B. Nicholl. 2012. Resveratrol enhances radiation sensitivity in prostate cancer by inhibiting cell proliferation and promoting cell senescence and apoptosis. *Cancer Science* n/a-n/a. [[CrossRef](#)]
11. Sumeyya Akyol, Zeynep Ginis, Ferah Armutcu, Gulfer Ozturk, M. Ramazan Yigitoglu, Omer Akyol. 2012. The potential usage of caffeic acid phenethyl ester (CAPE) against chemotherapy-induced and radiotherapy-induced toxicity. *Cell Biochemistry and Function* n/a-n/a. [[CrossRef](#)]
12. Fabiana Casanova, Julia Quarti, Danielly Cristiny Ferraz da Costa, Caroline Araújo Ramos, Jerson Lima da Silva, Eliane Fialho. 2012. Resveratrol chemosensitizes breast cancer cells to melphalan by cell cycle arrest. *Journal of Cellular Biochemistry* n/a-n/a. [[CrossRef](#)]
13. Yintao Ye, Wenqing Xu, Wei Zhong, Yajing Li, Chen Wang. 2011. Combination treatment with dihydrotanshinone I and irradiation enhances apoptotic effects in human cervical cancer by HPV E6 down-regulation and caspases activation. *Molecular and Cellular Biochemistry* . [[CrossRef](#)]
14. Miguel Asensi, Angel Ortega, Salvador Mena, Fatima Feddi, José M. Estrela. 2011. Natural polyphenols in cancer therapy. *Critical Reviews in Clinical Laboratory Sciences* 1-20. [[CrossRef](#)]
15. H. Yin, R. Guo, Y. Xu, Y. Zheng, Z. Hou, X. Dai, Z. Zhang, D. Zheng, H. Xu. 2011. Synergistic antitumor efficiency of docetaxel and curcumin against lung cancer. *Acta Biochimica et Biophysica Sinica* . [[CrossRef](#)]
16. Bandugula Venkata Reddy, N. Rajendra Prasad. 2011. 2-deoxy-D-glucose combined with ferulic acid enhances radiation response in non-small cell lung carcinoma cells. *Central European Journal of Biology* **6**:5, 743-755. [[CrossRef](#)]
17. Geumho Lee, Tae Won Choi, Chulwon Kim, Dongwoo Nam, Seok-Geun Lee, Hyeung-Jin Jang, Jun-Hee Lee, Jae-Young Um, Sang Hoon Jung, Bum Sang Shim, Kyoo Seok Ahn, Kwang Seok Ahn. 2011. Anti-inflammatory activities of Reynoutria elliptica through suppression of mitogen-activated protein kinases and nuclear factor- $\kappa$ B activation pathways. *Immunopharmacology and Immunotoxicology* 1-11. [[CrossRef](#)]
18. Ensaf M. Al-Hujaily, Ameera Gaafar Mohamed, Ibtehaj Al-Sharif, Khairia M. Youssef, Pulicat S. Manogaran, Basem Al-Otaibi, Amal Al-Haza'a, Ibrahim Al-Jammaz, Khaled Al-Hussein, Abdelilah Aboussekhra. 2011. PAC, a novel curcumin

- analogue, has anti-breast cancer properties with higher efficiency on ER-negative cells. *Breast Cancer Research and Treatment* **128**:1, 97-107. [[CrossRef](#)]
19. Neetu Singh, Manisha Nigam, Vishal Ranjan, Deeba Zaidi, Vivek Kumar Garg, Sharad Sharma, Rashmi Chaturvedi, Rishi Shankar, Sadan Kumar, Ramesh Sharma, Kalyan Mitra, Anil K. Balapure, Srikanta K. Rath. 2011. Resveratrol as an adjunct therapy in cyclophosphamide-treated MCF-7 cells and breast tumor explants. *Cancer Science* **102**:5, 1059-1067. [[CrossRef](#)]
  20. Subburayan Karthikeyan, Govindhasamy Kanimozhi, Nagarajan Rajendra Prasad, Rajendran Mahalakshmi. 2011. Radiosensitizing effect of ferulic acid on human cervical carcinoma cells in vitro. *Toxicology in Vitro* . [[CrossRef](#)]
  21. Tamer Refaat, Amr Elsaid, Nashaat Lotfy, Krystyna Kiel, William Small, Phillip Nickers, Eric Lartigau. 2011. Concomitant chemoradiotherapy with high dose rate brachytherapy as a definitive treatment modality for locally advanced cervical cancer. *Alexandria Journal of Medicine* **47**:1, 15-24. [[CrossRef](#)]
  22. Yongping Liu, Yang Ling, Wenjing Hu, Li Xie, Lixia Yu, Xiaoping Qian, Binxia Zhang, Baorui Liu. 2011. The Herb Medicine Formula “Chong Lou Fu Fang” Increases the Cytotoxicity of Chemotherapeutic Agents and Down-Regulates the Expression of Chemotherapeutic Agent Resistance-Related Genes in Human Gastric Cancer Cells In Vitro. *Evidence-Based Complementary and Alternative Medicine* **2011**, 1-10. [[CrossRef](#)]
  23. N. M. Rogers, S. Kireta, P. T. H. Coates. 2010. Curcumin induces maturation-arrested dendritic cells that expand regulatory T cells in vitro and in vivo. *Clinical & Experimental Immunology* **162**:3, 460-473. [[CrossRef](#)]
  24. Yolanda Sánchez, Donna Amrán, Elena de Blas, Patricio Aller. 2010. Arsenic trioxide as an anti-tumour agent: mechanisms of action and strategies of sensitization. *Journal of Applied Biomedicine* **8**:4, 199-208. [[CrossRef](#)]
  25. Amit K. Garg, Anuja Jhingran, Ann H. Klopp, Bharat B. Aggarwal, Ajai B. Kunnumakkara, Russell R. Broadus, Patricia J. Eifel, Thomas A. Buchholz. 2010. Expression of Nuclear Transcription Factor Kappa B in Locally Advanced Human Cervical Cancer Treated With Definitive Chemoradiation. *International Journal of Radiation OncologyBiologyPhysics* **78**:5, 1331-1336. [[CrossRef](#)]
  26. Wei Liu, Lan Tang, Ling Ye, Zheng Cai, Bijun Xia, Jiajie Zhang, Ming Hu, Zhongqiu Liu. 2010. Species and Gender Differences Affect the Metabolism of Emodin via Glucuronidation. *The AAPS Journal* **12**:3, 424-436. [[CrossRef](#)]
  27. Guillaume Jacquemin, Sarah Shirley, Olivier Micheau. 2010. Combining naturally occurring polyphenols with TNF-related apoptosis-inducing ligand: a promising approach to kill resistant cancer cells?. *Cellular and Molecular Life Sciences* **67**:18, 3115-3130. [[CrossRef](#)]
  28. Bunliang Suphim, Auemduan Prawan, Upa Kukongviriyapan, Sarinya Kongpetch, Benjaporn Buranrat, Veerapol Kukongviriyapan. 2010. Redox modulation and human bile duct cancer inhibition by curcumin. *Food and Chemical Toxicology* **48**:8-9, 2265-2272. [[CrossRef](#)]
  29. Jia-Rong Fan, Ting-Hsiang Huang, Chen-Yu Wen, Tang-Long Shen, Tsai-Kun Li. 2010. Sodium salicylate acts through direct inhibition of phosphoinositide 3-kinase-like kinases to modulate topoisomerase-mediated DNA damage responses. *European Journal of Pharmacology* **638**:1-3, 13-20. [[CrossRef](#)]
  30. Grégory Gatouillat, Emilie Balasse, Débora Joseph-Pietras, Hamid Morjani, Claudie Madoulet. 2010. Resveratrol induces cell-cycle disruption and apoptosis in chemoresistant B16 melanoma. *Journal of Cellular Biochemistry* **110**:4, 893-902. [[CrossRef](#)]
  31. K. C. Nicolaou, Qiang Kang, T. Robert Wu, Chek Shik Lim, David Y.-K. Chen. 2010. Total Synthesis and Biological Evaluation of the Resveratrol-Derived Polyphenol Natural Products Hopeanol and Hopeahainol A. *Journal of the American Chemical Society* **132**:21, 7540-7548. [[CrossRef](#)]
  32. James C. Lee, Paul A. Kinniry, Evguenia Arguiri, Matthew Serota, Stathis Kanterakis, Shampa Chatterjee, Charalambos C. Solomides, Prashanthi Javvadi, Constantinos Koumenis, Keith A. Cengel, Melpo Christofidou-Solomidou. 2010. Dietary Curcumin Increases Antioxidant Defenses in Lung, Ameliorates Radiation-Induced Pulmonary Fibrosis, and Improves Survival in Mice. *Radiation Research* **173**:5, 590-601. [[CrossRef](#)]
  33. Atsushi Sato, Kaori Sakurada, Toshihiro Kumabe, Toshio Sasajima, Takaaki Beppu, Kenichiro Asano, Hiroki Ohkuma, Akira Ogawa, Kazuo Mizoi, Teiji Tominaga, Chifumi Kitanaka, Takamasa Kayama. 2010. Association of stem cell marker CD133 expression with dissemination of glioblastomas. *Neurosurgical Review* **33**:2, 175-184. [[CrossRef](#)]
  34. Christina Eder-Czembirek, Boban M. Erovic, Cornelia Czembirek, Markus Brunner, Edgar Selzer, Richard Pötter, Dietmar Thurnher. 2010. Betulinic Acid a Radiosensitizer in Head and Neck Squamous Cell Carcinoma Cell Lines. *Strahlentherapie und Onkologie* **186**:3, 143-148. [[CrossRef](#)]
  35. Marcello Iriti, Franco Faoro Bioactive Chemicals and Health Benefits of Grapevine Products 581-620. [[CrossRef](#)]



36. Meng Xu, Liang-He Sheng, Xi-Hai Zhu, Shi-Bin Zeng, Guo-Jun Zhang. 2010. Reversal Effect of Stephania Tetrandra-Containing Chinese Herb Formula SENL on Multidrug Resistance in Lung Cancer Cell Line SW1573/2R120. *The American Journal of Chinese Medicine* **38**:02, 401-413. [[CrossRef](#)]
37. Rakesh Madhusoodhanan, Mohan Natarajan, Jamunarani Veeraraghavan Nisha Singh, Ambarish Jamgade, Vibhudutta Awasthi, Shrikant Anant, Terence S. Herman, Natarajan Aravindan. 2009. Effect of Black Raspberry Extract in Inhibiting NF- $\kappa$ B Dependent Radioprotection in Human Breast Cancer Cells. *Nutrition and Cancer* **62**:1, 93-104. [[CrossRef](#)]
38. Yolanda Sánchez, Consuelo Calle, Elena de Blas, Patricio Aller. 2009. Modulation of arsenic trioxide-induced apoptosis by genistein and functionally related agents in U937 human leukaemia cells. Regulation by ROS and mitogen-activated protein kinases. *Chemico-Biological Interactions* **182**:1, 37-44. [[CrossRef](#)]
39. 2009. Combined Treatment of Nonsteroidal Anti-inflammatory Drugs and Genistein Synergistically Induces Apoptosis via Induction of NAG-1 in Human Lung Adenocarcinoma A549 Cells. *Journal of Life Science* **19**:8, 1073-1080. [[CrossRef](#)]
40. Hui He, Laurie Chen, Ming Zhai, John Z. S. Chen. 2009. Genistein down-regulates the constitutive activation of nuclear factor- $\kappa$ B in human multiple myeloma cells, leading to suppression of proliferation and induction of apoptosis. *Phytotherapy Research* **23**:6, 868-873. [[CrossRef](#)]
41. P.J. Wills, V.V. Asha. 2009. Chemopreventive action of Lygodium flexuosum extract in human hepatoma PLC/PRF/5 and Hep 3B cells. *Journal of Ethnopharmacology* **122**:2, 294-303. [[CrossRef](#)]
42. Nathan R. Perron, Julia L. Brumaghim. 2009. A Review of the Antioxidant Mechanisms of Polyphenol Compounds Related to Iron Binding. *Cell Biochemistry and Biophysics* **53**:2, 75-100. [[CrossRef](#)]
43. Bharat B. Aggarwal, Bokyoung Sung. 2009. Pharmacological basis for the role of curcumin in chronic diseases: an age-old spice with modern targets. *Trends in Pharmacological Sciences* **30**:2, 85-94. [[CrossRef](#)]
44. Reo Iwasaki, Kinji Ito, Takaomi Ishida, Makoto Hamanoue, Souichi Adachi, Toshiki Watanabe, Yuko Sato. 2009. Catechin, green tea component, causes caspase-independent necrosis-like cell death in chronic myelogenous leukemia. *Cancer Science* **100**:2, 349-356. [[CrossRef](#)]
45. Maha H. Elamin, Zakia Shinwari, Siti-Faujiah Hendrayani, Hindi Al-Hindi, Essam Al-Shail, Yasser khafaga, Amani Al-kofide, Abdelilah Aboussekhra. 2009. Curcumin inhibits the Sonic Hedgehog signaling pathway and triggers apoptosis in medulloblastoma cells. *Molecular Carcinogenesis* n/a-n/a. [[CrossRef](#)]
46. Majda Hadolin Kolar, Simona Urban*et al*, Duřanka Dimitrijevi*et al* Nature Knows Best: Where Nature and Beauty Meet 399-419. [[CrossRef](#)]
47. Petras Juzenas, Wei Chen, Ya-Ping Sun, Manuel Alvaro Neto Coelho, Roman Generalov, Natalia Generalova, Ingeborg Lie Christensen. 2008. Quantum dots and nanoparticles for photodynamic and radiation therapies of cancer. *Advanced Drug Delivery Reviews* **60**:15, 1600-1614. [[CrossRef](#)]
48. Joydeb Kumar Kundu, Young-Joon Surh. 2008. Cancer chemopreventive and therapeutic potential of resveratrol: Mechanistic perspectives. *Cancer Letters* **269**:2, 243-261. [[CrossRef](#)]
49. M HEINRICH, J PRIETO. 2008. Diet and healthy ageing 2100: Will we globalise local knowledge systems?. *Ageing Research Reviews* **7**:3, 249-274. [[CrossRef](#)]
50. Isabel Villegas, Susana Sánchez-Fidalgo, Catalina Alarcón de la Lastra. 2008. New mechanisms and therapeutic potential of curcumin for colorectal cancer. *Molecular Nutrition & Food Research* **52**:9, 1040-1061. [[CrossRef](#)]
51. Hui He, Laurie Chen, Ming Zhai, John Z.S. Chen. 2008. Genistein down-regulates the constitutive activation of nuclear factor- $\kappa$ B of bone marrow stromal cells in multiple myeloma, leading to suppression of gene expression and proliferation. *Drug Development Research* **69**:4, 219-225. [[CrossRef](#)]
52. Miguel López-Lázaro. 2008. Anticancer and carcinogenic properties of curcumin: Considerations for its clinical development as a cancer chemopreventive and chemotherapeutic agent. *Molecular Nutrition & Food Research* . [[CrossRef](#)]
53. B. Sung, M. K. Pandey, K. S. Ahn, T. Yi, M. M. Chaturvedi, M. Liu, B. B. Aggarwal. 2008. Anacardic acid (6-nonadecyl salicylic acid), an inhibitor of histone acetyltransferase, suppresses expression of nuclear factor- $\kappa$ B-regulated gene products involved in cell survival, proliferation, invasion, and inflammation through inhibition of the inhibitory subunit of nuclear factor- $\kappa$ B kinase, leading to potentiation of apoptosis. *Blood* **111**:10, 4880-4891. [[CrossRef](#)]
54. Richard A Baxter. 2008. Anti-aging properties of resveratrol: review and report of a potent new antioxidant skin care formulation. *Journal of Cosmetic Dermatology* **7**:1, 2-7. [[CrossRef](#)]
55. G. Filomeni, I. Graziani, G. Rotilio, M. R. Ciriolo. 2007. trans-Resveratrol induces apoptosis in human breast cancer cells MCF-7 by the activation of MAP kinases pathways. *Genes & Nutrition* **2**:3, 295-305. [[CrossRef](#)]

56. Amit Deorukhkar, Sunil Krishnan, Gautam Sethi, Bharat B Aggarwal. 2007. Back to basics: how natural products can provide the basis for new therapeutics. *Expert Opinion on Investigational Drugs* **16**:11, 1753-1773. [[CrossRef](#)]
57. Binod Kumar, Jayashree Joshi, Amit Kumar, Badri N. Pandey, Banasri Hazra, Kaushala P. Mishra. 2007. Radiosensitization by diospyrin diethylether in MCF-7 breast carcinoma cell line. *Molecular and Cellular Biochemistry* **304**:1-2, 287-296. [[CrossRef](#)]
58. Sharmila Shankar, Imtiaz Siddiqui, Rakesh K. Srivastava. 2007. Molecular mechanisms of resveratrol (3,4,5-trihydroxy-trans-stilbene) and its interaction with TNF-related apoptosis inducing ligand (TRAIL) in androgen-insensitive prostate cancer cells. *Molecular and Cellular Biochemistry* **304**:1-2, 273-285. [[CrossRef](#)]
59. B ANNABI, J CURRIE, A MOGHRABI, R BELIVEAU. 2007. Inhibition of HuR and MMP-9 expression in macrophage-differentiated HL-60 myeloid leukemia cells by green tea polyphenol EGCg. *Leukemia Research* **31**:9, 1277-1284. [[CrossRef](#)]
60. Jia Wei, Baorui Liu, Lifeng Wang, Xiaoping Qian, Yitao Ding, Lixia Yu. 2007. Synergistic interaction between tetrandrine and chemotherapeutic agents and influence of tetrandrine on chemotherapeutic agent-associated genes in human gastric cancer cell lines. *Cancer Chemotherapy and Pharmacology* **60**:5, 703-711. [[CrossRef](#)]
61. Shramana Mandal, Ashish Kumar Mandal. 2007. Malignant fibrous histiocytoma following radiation therapy and chemotherapy for Hodgkin's lymphoma. *International Journal of Clinical Oncology* **12**:1, 52-55. [[CrossRef](#)]
62. Ganesh Chandra Jagetia, Bharat B. Aggarwal. 2007. "Spicing Up" of the Immune System by Curcumin. *Journal of Clinical Immunology* **27**:1, 19-35. [[CrossRef](#)]
63. Giuseppe Filomeni , Maria R. Ciriolo . 2006. Redox Control of Apoptosis: An Update. *Antioxidants & Redox Signaling* **8**:11-12, 2187-2192. [[Abstract](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
64. H Pelicano, D S Martin, R-H Xu, P Huang. 2006. Glycolysis inhibition for anticancer treatment. *Oncogene* **25**:34, 4633-4646. [[CrossRef](#)]